

Grape Polyphenols to Arrest *In Vitro* Proliferation of Human Leukemia Cells: A Systematic Review and Meta-analysis

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1

2 Abstract

3 Leukemia is a heterogeneous group of hemopoietic cancers, which accounts for
4 2.6% of new cases per year of total cancer incidence worldwide. Grapes and grape-
5 derived products, such as grape juice, are naturally rich in polyphenols, bioactive
6 compounds with antioxidant properties. Certain polyphenols have been proved to alter
7 oxidative balance, both in inducing apoptosis in cancer cells and in preventing cancer
8 development via controlling oxidative stress. To assess the therapeutic potential of grape
9 polyphenols in the treatment of leukemia, a systematic review and meta-analysis of the
10 reported data on leukemia was carried out. Following the PRISMA guide, a literature
11 review of published papers on leukemia and polyphenols from the last 50 years was
12 conducted, and 17 scientific articles published from 2002-2017 were included in the
13 study. Resveratrol 50 μm had the highest growth inhibition effect (67%) followed by
14 quercetin (30%). The results also point to a differential effect of polyphenols based on
15 cell lineage; monocytes- and myelocytic-derived cell lines are the most susceptible, with
16 a mean of 85% and 64% proliferation inhibition, respectively. Moreover, results show
17 that growth inhibition cannot be associated with a molecular effect of polyphenols on the
18 cell cycle arrest.

19

20

21 **Keywords:** grape polyphenols, leukemia, cell proliferation, meta-analysis

22

23

24 **Introduction**

25 Several interesting compounds with applications in the field of healthcare can be
26 found in vine, such as polyphenols. Polyphenols are a large family of secondary
27 metabolites that confer selective advantages against ecological stress and are extensively
28 distributed among fruits and vegetables ^[1]. Polyphenols are compounds that contain at
29 least one aromatic ring bearing one or more hydroxyl substituents (phenols) in their
30 chemical structure. They can be classified into groups according to the complexity of their
31 chemical structure and molecular weight ^[2-4].

32 Polyphenols present in grape berries are mainly concentrated in the skin and seeds
33 and constitute between 28-35% and 60-70%, respectively, of the total phenols that can be
34 extracted from grapes ^[5]. Regarding their structure, the most common polyphenol
35 families in grapes are phenolic acids (such as coumaric, ellagic, caffeic, or gallic),
36 flavonoids (including apigenin, catechin and epicatechin), flavanols (such as quercetin or
37 kaempferol) and stilbenes (which include resveratrol or pterostilbene) ^[6, 7]. These
38 substances have proved to be important natural antioxidant and anti-inflammatory
39 products that serve as free radical scavengers. As a result of this, grape berries and grape-
40 derivatives have become important compounds with applications in the healthcare and
41 food industries ^[8]. Several epidemiological studies have shown that a polyphenol-rich diet
42 prevents the development of neurodegenerative and cardiovascular diseases, cancer,
43 diabetes, allergy, and autoimmune disorders ^[9-11]. Many studies have been developed to
44 analyze the putative beneficial effects of the consumption of grapes, raisins, grape juice,
45 wine and grape extracts in the progression or prevention of specific human diseases ^{[12-}
46 ^{15]}. Similarly, purified polyphenols such as quercetin, resveratrol and others have been
47 tested in studies carried out *in vitro* and *in vivo* to assess their therapeutic use in the
48 treatment of cancer and other chronic diseases ^[16, 17].

49 The molecular mechanisms by which polyphenols act in the cells are not yet well
50 known. Nevertheless, some studies have shown that the protective and therapeutic effect
51 of polyphenols is carried out through different metabolic, cellular and molecular
52 mechanisms that modulate the gene expression, signaling pathways and epigenetic
53 changes in the cells ^[18,19]. It has been demonstrated that many polyphenols induce
54 apoptosis specifically in pathological cells ^[20,21], which has resulted in several studies
55 testing polyphenols from different natural sources as chemotherapeutic agents against
56 cancer ^[17, 28, 29].

57 Leukemia is a heterogeneous disease resulting from the transformation of
58 hematopoietic stem or progenitor cells, and it is the 10th most prevalent cause of cancer
59 in the world ^[1]. In the US, from 2009-2015, the number of new cases of leukemia per year
60 was 14.1 per 100,000 inhabitants, and the number of deaths per 100,000 inhabitants was
61 6.5 ^[27]. In Spain, 3,419 people died in 2016 due to leukemia, and the incidence of the
62 disease in 2017 was 5,424 cases. There are different types of leukemia that can be divided
63 into acute or chronic in function of the maturity stage of the malignant cells, and into
64 lymphoid or myeloid in relation to the cell lineage. Based on malignant transformation
65 experienced by cell lineages and their clinical characteristics, the main types of leukemia
66 are acute myeloid leukemia (AML), chronic myeloid leukemia (CML), acute lymphocytic
67 leukemia (ALL) and chronic lymphocytic leukemia (CLL) ^[28, 29]. Chronic types of
68 leukemia present higher incidence in adults; however, acute lymphocytic leukemia has
69 become the leading cause of death in children suffering from cancer ^[31]. Moreover, acute
70 leukemia patients usually require prolonged periods of hospitalization and their quality
71 of life is limited. In the US, incidence rates of acute leukemia (one of the main causes of
72 leukemia morbidity) equaled 5 cases per 100,000 inhabitants in 2018; in 2035, a total of
73 315,413 cases are expected ^[27].

74 Accordingly, many studies are currently being carried out to identify effective
75 treatments against leukemia. Several studies conducted in recent decades have shown that
76 grape pomace and seed extracts, rich in polyphenols, induce apoptosis and inhibit the
77 growth of leukemia cells ^[31,32]. However, the natural polyphenol more frequently used in
78 studies with leukemia cells is resveratrol ^[33]. There is evidence that resveratrol induces
79 apoptosis in the four main types of leukemia (AML, CML, ALL and CLL), but other
80 mechanisms, such as autophagy induction, cell cycle arrest or growth inhibition, are also
81 included in the molecular mechanisms induced by resveratrol in specific cell lines ^{[23, 34-}
82 ^{37]}. However, to date, no studies have been carried out to integrate all these results and
83 evaluate the putative therapeutic effect of grape polyphenols, as well as their potential use
84 as adjuvants in the treatment of leukemia patients. This could be of great interest for the
85 nutraceutical and food industries, given that grape-derived products have numerous
86 applications in dietary foods and supplements, especially in infant and sports products.

87 Thus, the aim of this study is to evaluate, based on the available literature, whether
88 grape polyphenols have the same antiproliferative effect on leukemia tumor lines
89 regardless of cell lineage and the type of polyphenol. For this purpose, a systematic
90 review and meta-analysis of research published from 2002-2017 has been carried out.

91

92 **Materials and Methods**

93 *Literature Search*

94 The literature search was conducted following the guidelines included in the
95 Preferred Reporting Items for Systematic Reviews and Meta-Analyses guide (PRISMA)
96 ^[38-40]. Accordingly, a systematic search was carried out using the following databases:
97 the Cochrane library ^[41], EMBASE ^[42], MEDLINE ^[43], CINAHL ^[44]. Scientific reports

98 included in the study were obtained using the following search terms: “(grape) AND
99 (polyphenol) AND (leukemia)”.

100

101 ***Study Eligibility Criteria***

102 The following inclusion criteria were used for screening: (1) controlled trials,
103 which refers to reports with control assays in every experiment, (2) studies conducted
104 with leukemia tumor cells or leukemia cell lines, and (3) studies using mixtures of
105 polyphenols from grape extracts or purified polyphenols present in grape berries.

106 Exclusion criteria were: (1) uncontrolled trials and (2) use of polyphenols from sources
107 other than grapes.

108

109 ***Study Selection***

110 Both the titles and abstracts of articles that met the criteria were independently
111 screened by two authors of the present study using Covidence Software (Covidence
112 systematic review software, Veritas Health Innovation, Melbourne, Australia).

113 Following this, two authors acted as reviewers by independently assessing full-text
114 articles of potentially eligible studies. In the case of a lack of consensus between the two
115 reviewers involved in the screening or in the assessment of the full-texts, authors
116 discussed the conflict and reached an agreement. We documented the reasons for the
117 exclusion of the full-text articles we assessed.

118

119 ***Quality Assessment***

120 Studies included in the meta-analysis were carried out with established leukemia
121 cell lines cultured *in vitro*. Cell proliferation was analyzed in the presence or absence of
122 specific polyphenols. In this type of study, it is not possible to analyze bias following

123 the methodology proposed in the Cochrane manual ^[45], as it is specific to the evaluation
124 of clinical studies. Nevertheless, all studies included encompass reliable selection
125 criteria and there is no risk of bias present. Furthermore, leukemia cell lines had been
126 described, and reagents used were purchased from commercial companies with certified
127 reliability such as Sigma-Aldrich, JF-Natural Technology Co. or ActiVin.

128

129 ***Data Extraction***

130 Data extraction from the selected studies was performed using the standardized
131 procedure developed by PRISMA ^[39, 40] including the following information: (1)
132 complete author name, (2) year of publication, (3) polyphenol name, (4) concentration
133 of polyphenol, (5) leukemia cell type, and (6) effect on cell proliferation. To perform
134 the sifting, Covidence ^[46] software was chosen since it facilitates working in
135 collaboration with Cochrane in order to improve the production and use of systematic
136 reviews for healthcare and well-being.

137

138 ***Statistical Analysis***

139 The purpose of this meta-analysis was to analyze the positive or negative effect of
140 grape polyphenols on the proliferation rate of human leukemia cell lines' growth *in*
141 *vitro*. Data on the effects caused by each polyphenol in the proliferation rate of the
142 different leukemia cell lines were treated separately. The evaluation of the effect of the
143 different polyphenols on cell proliferation, as well as the graphic representation of
144 results was performed using Review Manager 5.3 ^[47] software. A 95% confidence
145 interval (CI) was established for all outcomes, including cell proliferation and stage of
146 cell cycle arrest. The two main methods used to compare data of a study with
147 dichotomous variables are: analyzing the risk ratio (RR) or comparing the odds ratio

148 (OR) ^[48, 49]. The OR analysis is the recommended method to evaluate dichotomous
149 variables, but as the relative risk was higher than 10% in the present study, RR analysis
150 was used. Under these conditions, the OR analysis overestimates the relative risk
151 because the random frequency of the effect of the evaluated compound is not correctly
152 handled, and false positives or negatives can be added to the study, which leads to a
153 biased final result ^[48]. The RR analysis ^[50] was therefore used for risk estimation. Data
154 heterogeneity was measured by the chi-square (χ^2) test and the I^2 statistic. The level of
155 significance was defined as $p < 0.10$. The I^2 statistic represents the total variation due to
156 the heterogeneity, and I^2 values higher than 50% indicate significant heterogeneity ^[50].
157 A random effect (RE), instead of a fixed effect (FE), was used for the analysis because
158 the effect of polyphenols can change depending on the compound used and leukemia
159 cells treated. The fixed parameters in the study were the concentration of each
160 polyphenol and the leukemia cell lines used.

161

162 **Results**

163 The flow chart describing the process of selecting publications to be included in
164 the study is presented in Figure 1. Once duplicate results were removed, 132 articles
165 were selected as a result of the search carried out. The Titles and Abstracts of these
166 articles were reviewed by the authors against the previously established eligibility
167 criteria. Accordingly, 74 articles were excluded from the study, as they did not fulfill
168 the criteria. The remaining 58 articles were screened by full text. Overall, 24 studies met
169 the criteria, but only 17 articles, which included quantitative data useful for the
170 statistical analysis, were selected to conduct the meta-analysis ^[31, 32, 37, 51-64].

171 [Figure 1 near here]

172

173 *Study Characteristics*

174 The above-mentioned 17 articles included in the study were thoroughly analyzed
175 to obtain quantitative data regarding the viability of leukemia cells based on the type
176 and concentration of polyphenols added to the culture media. Culture conditions for the
177 different cell types used in the studies were the same: cells were grown at 37°C in an
178 incubator with 5% CO₂ atmosphere using an RPMI-1640 medium supplemented with
179 10% (v/v) fetal calf serum, L-glutamine, penicillin and streptomycin. The experimental
180 procedures to determine the effect of polyphenols in cell viability and proliferation were
181 carried out in a very similar way by different researchers. When cultured cells reached a
182 cell density of 10⁴-10⁶ cells/mL, polyphenols were added to the culture media and
183 incubated for 24-72 hours before measuring cell viability.

184 *Outcomes of Interest*

185 The three outcomes pursued in the present study were:

- 186 1. Analyze the effect of specific grape polyphenols in cell proliferation of
187 leukemia cells cultured *in vitro*.
- 188 2. Determine whether the effect of grape polyphenols in leukemia cell
189 proliferation depends on the type of leukemia cell line.
- 190 3. Determine whether cell cycle arrest is the molecular target of grape
191 polyphenols on leukemia cells.

192 Table 1 summarizes the included articles and the information extracted to evaluate the
193 three outcomes stated above.

194 [Table 1 near here]

195 *Polyphenols Analyzed*

196 There was a large variability in the type of polyphenols included in the articles
197 selected for the meta-analysis; 13 pure polyphenolic compounds and grape polyphenol

198 extract (GPE) were analyzed. The data included in the meta-analysis were obtained 48
199 hours after treatment with polyphenols at fixed concentrations: 50 μ M resveratrol,
200 apigenin, cyanidin glucoside, delphinidin, ellagic acid, kaempferol, malvidin, myricetin,
201 petunidin and pterostilbene, 25 μ M viniferin and vineatrol, 10 μ M quercetin and 50/100
202 μ g/mL GPE. It must be noted that the origin and polyphenol composition of the extracts
203 used in the studies were different, which is the reason why interventions with GPE are
204 always analyzed in a different subgroup. Espino et al.^[32] used Tempranillo grape seed
205 extracts at 6,160.01 \pm 160.14 mg/L gallic acid equivalents (GAE); León-González et
206 al.^[31] used grape pomace extracts at 365 mg/g GAE; Sharif et al.^[62] used red wine dry
207 powder extracts at 471 mg/g GAE; Mertens-Talcott et al.^[63] used Muscadine and
208 Sauvignon wine extracts to 10,269 \pm 36 and 9,932 \pm 22 mg/L GAE, respectively; Wang et
209 al.^[64], Gao et al.^[54] and Hong et al.^[57] used commercial grape seed extracts enriched
210 with proanthocyanidins, but there was no reference to GAE. The viable cells (called
211 “events” in figures 2, 3 and 4) following treatment were measured using different
212 methods. However, direct quantification using flow cytometry was the most common.

213 *Cell Lines Used in the Studies*

214 In relation to leukemia cell lines included in the analyzed studies, four of them
215 originated from early myeloid cells (Em): K562, HL60, AML14.3D10, KCL22; two
216 from monocytes (Mc): U937 and THP1; four from B lymphocytes (LB): WSU-CLL,
217 ESKOL, NALM-6, 232B4, and two from T lymphocytes (LT): MOLT-4 and JURKAT.
218 Moreover, HL60, U937, THP1 and AML14.3D10 are acute promyeloid and monocytic
219 leukemia cells, while K562 and KCL22 are chronic myeloid cell lines. In the
220 lymphocytic lineages, NALM-6, MOLT-4 and JURKAT are models of acute leukemia,
221 and 232B4, WSU-CLL and ESKOL, are chronic leukemia cells.

222 The number of experiments in the articles selected for meta-analysis that were
223 useful for the data extraction process is shown in Table 2, sorted by type of polyphenol
224 and cell line.

225 [Table 2 near here]

226

227 ***Outcome 1. The Effect of Specific Grape Polyphenols on the Proliferation of***
228 ***Leukemia Cell Lines***

229 Results obtained through this meta-analysis on the effect of grape polyphenols
230 on cell proliferation are presented in five different subgroups. Three of the subgroups
231 show the effects of resveratrol, quercetin and viniferin. The “others” subgroup clusters
232 the effect of other grape polyphenols (where only the results of one experiment are
233 available for each one), and the other subgroup shows the effect of GPE (Figure 2). It is
234 important to highlight that maximum heterogeneity (I^2) was obtained through statistical
235 analysis in all cases. This could be due to the different leukemia cell lines used at
236 variable cell concentrations across the different experiments and laboratories (Table 2).
237 In this case, risk ratio is employed as an indicator.

238 [Figure 2 near here]

239 As a preliminary conclusion, a positive overall effect can be considered
240 regarding the use of these chemical compounds as inhibitors of tumor line growth. This
241 can be seen through the presence of diamond-shaped symbols within every cluster to the
242 left side of the vertical line, showing the absence of a null effect in all cases.
243 Furthermore, following a deeper analysis, we infer that both quercetin and viniferin
244 cause a slight inhibitory effect, particularly when compared to resveratrol. The higher
245 number of studies including the use of resveratrol should be considered in the
246 discussion of these results. Similarly, a significant dispersion caused by the wide range

247 of cell lines used must also be considered. Upon comparing the results when using pure
248 polyphenols, resveratrol shows a high variability in the inhibition of cell proliferation
249 when different leukemia cells are analyzed. It must be noted that 13 interventions and
250 10 different cell lines have been assessed in the specific case of resveratrol, as opposed
251 to the 3 interventions and 3 cell lines evaluated for quercetin, and the 2 interventions
252 and 2 cell lines for viniferin. Nevertheless, results of the meta-analysis indicate that
253 resveratrol presents the greatest capacity to inhibit cell proliferation with a mean value
254 of 67% (95% CI, 56%-75%), which is significantly higher than the mean inhibition
255 found when quercetin (30%, 95% CI, 22%-38%) and viniferin (15%, 95% CI, 5%-23%)
256 are used. The subgroup “others”, which encompasses results of individual studies, was
257 evaluated using 10 different grape polyphenols (apigenin, myricetrin, delphinidin,
258 kaempferol, petunidin, malvidin, pterostilbene, cyanidin glucoside, vineatrol and ellagic
259 acid) and four leukemia cell lines. In this case, the average growth inhibition percentage
260 is 53% (95% CI, 40%-63%). However, the limitation of this subgroup, which includes
261 data from single experiments carried out with different polyphenols, must be taken into
262 account.

263 According to the number of studies meta-analyzed, 50 μ g/mL GPE was the
264 second most effective treatment. The average growth inhibition is 58% (95% CI, 46%-
265 67%), lower than those obtained with resveratrol (67%, 95% CI, 56%-75%). This is an
266 important point considering the GPE used in the studies have different origins and
267 polyphenol compositions; however, only six different cell lines were analyzed in
268 contrast to the ten lines used with resveratrol. No data coming from B-lymphocytes-
269 derived leukemia cells are available.

270 The results as a whole show the highest value of growth inhibition is the 67%
271 obtained upon exposure to 50 μ M resveratrol, followed by the 53% obtained with the

272 group of other polyphenols used at the same final concentration. It is true that when
273 leukemia cells are treated with GPE the percentage of growth inhibition is 58%, but in
274 these cases the polyphenol composition and concentration are unknown, so this value
275 can be used only as a reference, and not as an individual comparison.

276

277 ***Outcome 2. The Effect of Grape Polyphenols on the Proliferation of Human***

278 ***Leukemia Cells of Different Origin***

279 Results compiled in relation to outcome 2 refer to the influence of grape
280 polyphenols on the cell proliferation of leukemia cells from different lineages. The 12
281 cell lines used in the studies included in this meta-analysis can be subdivided into four
282 groups according to their source of origin: monocytes (U937 and THP1), early myeloid
283 cells (HL60, K562, KCL22 and AML14.3D10), B lymphocytes (WSU-CLL, ESKOL,
284 NALM-6 and 232B4) and T lymphocytes (MOLT-4 and JURKAT).

285 Studies performed with leukemia cell lines from lymphocytic origin represent
286 71% of the total and, as can be seen in Figure 3A, the average percentage of growth
287 inhibition due to the effect of pure polyphenols is 53% (95% CI, 41%-63%) and 44%
288 (95% CI, 28%-57%), for both T-and B-lymphocytic-derived leukemia cells,
289 respectively, which are lower than the values observed for early myeloid (64%, 95% CI,
290 49%-75%) and monocyte (85%, 95% CI, 65%-94%) derived-leukemia cells. When cells
291 were treated with pure polyphenols, the overall growth inhibition percentage obtained in
292 this outcome was 56% (95% CI, 47%-64%) (Figure 3A), similar to the 58% (95% CI,
293 46%-67%) obtained when cells were treated with GPE (Figure 3B).

294 Both cell lines derived from T lymphocytes were acute lymphoid leukemia
295 models, and it is necessary to highlight the effect of apigenin ^[53] and resveratrol ^[37] on
296 JURKAT cells, with growth inhibition percentages of 84% and 82%, respectively, as

297 well as the 92% growth inhibition demonstrated by pterostilbene^[63] on MOLT-4 cells
298 (Figure 3A). These data are the only ones misaligned with the trend followed by other
299 polyphenols^[53, 58, 61], which ranged between 7% and 42% growth inhibition. The mean
300 effect among all the polyphenols tested on this type of leukemia cells is 53% (95% CI,
301 41%-63%), similar to outcome 2's overall effect (56%, 95% CI, 47%-64%) (Figure 3A).
302 The average growth inhibition increased to 58% (95% CI, 37%-72%) when MOLT-4
303^[60] and JURKAT^[31, 37] cells were treated with GPE (Figure 3B).

304 [Figure 3 near here]

305 The subgroup of B-lymphocyte-derived leukemia cells includes data obtained
306 using four different polyphenols, three of which (except quercetin) belong to the
307 stilbene family. Four cell lines were also used, as most of the data was obtained from
308 chronic leukemia models (WSU-CLL, ESKOL, 232B4) and only one from the acute
309 NALM-6 leukemia cells. The average inhibition of proliferation in B-lymphocyte-
310 derived leukemia cells is 44% (95% CI, 28%-57%), indicating that these cells are less
311 sensitive to the effect of grape polyphenols than cell lines derived from T lymphocytes.
312 Importantly, resveratrol shows a 17% growth inhibition on NALM-6 cells^[55], while on
313 WSU-CLL, 232B4 and ESKOL cells the percentage of growth inhibition is 46%, 64%
314 and 67%, respectively^[31, 51, 56]. It was also noted that viniferin exerts little inhibition on
315 the growth of both ESKOL (19%) and WSU-CLL (10%) cells^[53].

316 The subgroup of early myeloid leukemia cells includes five studies carried out
317 with three different cell lines and resveratrol. It is worth highlighting the 88% growth
318 inhibition rate obtained with resveratrol in HL60 cells^[37], which is the only acute
319 myelocytic leukemia line included in this subgroup (Figure 3A). The mean inhibitory
320 effect of resveratrol on the growth of these cells is 64% (95% CI, 49%-75%), higher than
321 the 59% (95% CI, 33%-75%) obtained with GPE (Figure 3B). Nevertheless, the three

322 interventions with GPE were carried out by employing commercial polyphenol extracts
323 of unknown composition, thus making it difficult to draw reliable conclusions.

324 Finally, the subgroup of monocyte-derived cell lines includes experimental data
325 from a study carried out with resveratrol and two leukemia cell lines ^[37]. The result of
326 the meta-analysis indicates that these leukemia cells experience an average growth
327 inhibition of 85% (95% CI, 65%-94%), making them the most sensitive to the inhibitory
328 effect of pure grape polyphenols; precisely, to resveratrol (Figure 3A). Nevertheless,
329 only two values are included in this subgroup. The only study carried out on these cells
330 with GPE ^[54] shows a growth inhibition percentage of 47% (Figure 3B), much lower
331 than the effect of resveratrol.

332

333 ***Outcome 3. The Effect of Grape Polyphenols in Cell Cycle Arrest***

334 There was a large variability of targets and techniques used to assess cell death
335 and the growth inhibition of leukemia cells in the articles included in this meta-analysis.
336 Nevertheless, eight articles were detected that included quantitative data related to cell
337 cycle arrest in G₀ phase after the treatment of leukemia cells with grape polyphenols.
338 Therefore, the third outcome seeks to determine whether grape polyphenols inhibit
339 leukemia cell proliferation by arresting the cell cycle in G₀ phase. To achieve this, data
340 from experiments conducted with four grape polyphenols were used: resveratrol (50
341 μM, 24 hours), quercetin (10 μM, 48 hours), ellagic acid (10 μM, 48 hours), and
342 pterostilbene (44 μM, 48 hours). Data from five experiments performed using GPE (100
343 μg/mL, 24 hours) were also included.

344 The results obtained indicate that grape polyphenols are not directly involved in
345 cell cycle arrest of *in vitro* cultured leukemia cell lines (Figure 4). In the subgroup
346 including treatment with pure polyphenols, the mean effect observed was 1% (95% CI, -

347 0,7%-8%); thus, it can be concluded that the analyzed polyphenols affect cell viability
348 through a different mechanism than G₀ cell cycle arrest (Figure 4). Four out of six data
349 sets show no effect, and when pterostilbene was added to MOLT-4, the opposite effect
350 to what was expected occurred. Only one intervention carried out with quercetin on
351 MOLT-4 cells showed induction of cell cycle arrest. In addition, quercetin shows a
352 small positive effect on 232B4 cells, while resveratrol shows no effect on cell cycle
353 arrest either on 232B4 or on K562 cells.

354 The subgroup of the data obtained with GPE showed no effect on cell cycle arrest
355 on neither early myeloid ^[37, 64] nor on T-lymphocytes ^[60, 62] derived-leukemia cells
356 (Figure 4). The induction of cell cycle arrest in the G₀ phase was only observed once, in
357 the study carried out with HL60 cells treated with *Tempranillo* grape seed extracts at
358 6,160±160 mg/L GAE ^[32]. In contrast, the rest of the studies indicate the opposite effect,
359 stimulating the cell cycle progression.

360 [Figure 4 near here]

361 In conclusion, as can be seen in Figure 4, the dispersion of results regarding the
362 influence on the cell cycle is greater with GPE than with pure polyphenols. The
363 subgroup of pure polyphenols shows that 4 out of the 6 observations have a value of
364 almost 1, which implies that polyphenols influence neither the promotion nor the
365 impediment of cell-cycle arrest. In contrast, GPE shows that 3 of the 5 experiments
366 assessed are negative for cell cycle arrest. Moreover, the addition of these extracts does
367 not stop the cell cycle, but rather it encourages it.

368

369 Discussion

370 There are systematic reviews and meta-analysis studies that analyze the effect of
371 polyphenols in the prevention and treatment of some types of cancer, such as breast,

372 lung, prostatic or cervical cancer [24, 26, 65-68]. However, no studies related to leukemia
373 have been carried out. The majority of the studies support the positive effects of these
374 antioxidant compounds in the treatment of cancer; hence the interest in this study to
375 determine whether grape polyphenols have antiproliferative effects in leukemia. The
376 study has been conducted with experimental data on different human leukemia cell lines
377 cultured *in vitro*, given the lack of data in relevant clinical trials of leukemia patients
378 treated with polyphenols.

379 17 studies were included in the meta-analysis and the results obtained lead to the
380 conclusion that grape polyphenols have an antiproliferative effect on human leukemia
381 cell lines cultured *in vitro*, although this effect is dependent on the type of polyphenol
382 and cell line studied. It must be noted that results indicate there is a large heterogeneity
383 among the analyzed data, which can be explained by the type of studies evaluated in
384 which the cell concentration changed in each study, and by the variety of polyphenols
385 and cell lines used. Therefore, we consider the inability to perform a risk of bias
386 (because all tools available are not suitable for experimental studies like this) a
387 limitation of this study. On the other hand, despite all heterogeneity found in the
388 metanalyses, we could only explore it by subgrouping studies depending on the
389 concentration of each polyphenol and the leukemia cell lines used.

390 Outcome 1 indicates a global antiproliferative effect of grape polyphenols on
391 leukemia cells, with an average value of growth inhibition of 57% (95% CI, 49%-63%).
392 Among the pure polyphenols analyzed, resveratrol shows the greatest effect, with a 67%
393 (95%CI, 56%-75%) growth inhibition. This result agrees with previous reports
394 indicating that resveratrol could be a good bioactive compound for the prevention and
395 therapeutic use against different types of cancer [16, 28]. Quercetin and viniferin also
396 show a positive effect, but the mean growth inhibition is 30% (95% CI, 22%-38%) and

397 15% (95% CI, 5%-23%), respectively. It should be highlighted that the number of
398 studies including resveratrol is much higher than the studies carried out with other
399 polyphenols. However, it is noteworthy that the overall effect of the subgroup that
400 includes other grape polyphenols results in a 53% (95% CI, 40%-63%) inhibition of
401 growth. This subgroup includes data coming from single experiments carried out with
402 different polyphenols on leukemia cell lines of various lineages, so the variability of the
403 effect is high. It is interesting to highlight the effect of apigenin (flavonoid) and
404 pterostilbene (stilbene) on JURKAT and MOLT-4 cells, both acute T-lymphoblastic-
405 leukemia lines, that inhibit cell growth by 84% and 92%, respectively. Apigenin has
406 been proposed as a potential natural anti-cancer agent promoting apoptosis through
407 activation of extrinsic caspase-dependent pathways in cancer cells from different origins
408 [69-71]. Moreover, *in vivo* studies carried out with mouse models of certain cancers
409 showed a potential therapeutic use of apigenin at 25 mg /Kg [70]. Pterostilbene belongs
410 to the stilbene family, of which resveratrol is the main representative, and it has been
411 demonstrated that these compounds interfere with metabolic pathways directly linked to
412 senescence and induction of apoptosis in tumor cells [16, 72-74].

413 The growth inhibition observed when treating leukemia cells with GPE is 58%
414 (95% CI, 46%-67%), higher than the value observed using other polyphenols (53%,
415 95% CI, 40%-63%), but lower than the 67% (95% CI, 56%-75%) obtained with 50 μ M
416 resveratrol. The interpretation of these data is difficult because the specific polyphenol
417 composition of GPE is unknown. However, these results would support the hypothesis
418 that a combination of natural polyphenols rather than a single compound may be more
419 beneficial and effective in the treatment and prevention of leukemia [8].

420 The results of the effect of grape polyphenols on leukemia cells grouped by
421 cellular lineage (outcome 2) indicate an antiproliferative effect of these compounds,

422 regardless of the cellular origin. The mean value of growth inhibition is 56% (95% CI,
423 47%-64%) with pure polyphenols and 58% (95% CI, 37%-72%) using GPE.

424 The subgroup of the monocyte-derived cells shows the highest percentage of
425 growth inhibition at 85% (95% CI, 65%-94%), followed by the early myeloid-derived
426 cells at 64% (95% CI, 49%-75%). In all of them, the treatment was carried out with 50
427 μ M resveratrol, and the highest inhibitory effect is shown in the acute leukemia cells
428 from monocytic (TPH1, U937) and promyelocytic origin (HL60). The chronic myeloid
429 leukemia cells KCL22 and K562 show more variability and lower inhibition
430 percentages (21%-69%). The few data available with GPE rich in proanthocyanidins
431 show variable percentages of growth inhibition and different behaviors (47% in U937
432 cells and 80% in K562). Nevertheless, there is evidence that demonstrates the
433 antiproliferative effect of proanthocyanidins, either alone or in combination with other
434 polyphenols like resveratrol in colorectal, prostate and breast cancer cells ^[75-78].

435 The leukemia cells of lymphocytic lineages showed the lowest inhibitory effects.
436 The largest number of studies was carried out with T lymphocyte cell lines and the
437 average growth inhibition was 53% (95% CI, 41%-63%), whilst in the cell lines derived
438 from B lymphocytes a 44% (95% CI, 28%-57%) growth inhibition rate was obtained.
439 This result indicates that B-lymphocyte-derived cells are less sensitive to the
440 antiproliferative effect induced by grape polyphenols. Importantly, there are three
441 polyphenols that produce outstanding values of growth inhibition on T-lymphocyte-
442 derived cells; apigenin on JURKAT and the stilbenes, and resveratrol and pterostilbene
443 on MOLT-4.

444 The results allow us to affirm that the antiproliferative effect of grape polyphenols
445 in leukemia cell lines derived from early myeloid cells and monocytes is higher than in
446 cells derived from T and B lymphocytes. Moreover, resveratrol has a higher effect in the

447 acute myeloid- and monocyte-derived cells than in the chronic cell lines. This is a very
448 important aspect to consider for the development of future *in vitro* and *in vivo*
449 experiments. The molecular differences in the intracellular metabolic and signaling
450 pathways in acute and chronic leukemia cells can be directly involved in the
451 effectiveness of the treatment with resveratrol or other polyphenols.

452 Outcome 3 indicates that polyphenols have heterogeneous effects regarding the
453 cell cycle arrest. Only interventions with quercetin can induce the arrest of the cell cycle
454 in B- and T-derived-leukemia cells. In contrast, resveratrol and ellagic acid show no
455 effect, and the treatment with pterostilbene promotes the cell cycle. When cells were
456 treated with GPE, four out of five observations show no effect. These results indicate
457 that the growth inhibition promoted by grape polyphenols on leukemia cells is not
458 produced by arresting the cell cycle in the G₀ phase.

459 The overall conclusion of the meta-analysis is that grape polyphenols have a
460 detrimental effect in the growth of leukemia cells *in vitro*. The degree or magnitude of
461 this effect depends on the source of the cell lines and the specific polyphenol or GPE
462 used. Recent studies have shown an antiangiogenic activity of polyphenol extracts on
463 cancer cells [22] and the ability to modulate epigenetic changes directly involved in
464 cancer development [78, 79]. More specifically, resveratrol has been proved to modulate
465 autophagy and induce apoptosis in leukemia cells [23, 35, 36]. Furthermore, studies with
466 animal models lead to propose that the consumption of polyphenols in childhood and
467 even in the maternal diet could prevent the development of chronic diseases in
468 adulthood [18]. These data, together with the results obtained through this meta-analysis
469 showing the antiproliferative effect of grape polyphenols (mainly resveratrol) on
470 leukemia cells, support the development of *in vivo* studies to assess their usefulness in
471 the treatment and prevention of leukemia.

472 However, it must be noted that the concentration of polyphenols used in the *in*
473 *vitro* assays is almost 25 times higher than the average estimated concentration of
474 resveratrol present in grape juice ^[16]. Moreover, the bioavailability and
475 pharmacokinetics of polyphenols mainly depend on the doses ingested, the food matrix
476 and the gut microbiota. In the case of resveratrol, a safe and efficient dose has been
477 proved between 1g and 5 g per day, although adverse effects can be detected in some
478 individuals ^[88]. There are studies working on new formulations to enhance the
479 bioavailability of resveratrol, mostly by means of increasing its hydrophilicity, stability
480 and bioaccessibility. Studies carried out with resveratrol and pterostilbene in animal
481 models showed that both can be rapidly absorbed and metabolized in their
482 glucuronidated and sulfated forms. Moreover, due to its chemical structure of a
483 dimethyl ether analog of resveratrol, pterostilbene showed a better membrane
484 permeability and metabolic stability, which consequently increases its bioavailability
485 and pharmacokinetic properties ^[92]. In a recent study, polyphenolic derivatives with
486 increased lipophilicity have shown better cellular absorption, greater antioxidant
487 capacity and greater antiproliferative effect on tumor cells grown *in vitro* ^[94]. Also, new
488 procedures to protect polyphenols from digestive degradation have been studied, in
489 order to allow these bioactive compounds to reach their therapeutic targets through food
490 ^[94]. However, it is important to highlight that resveratrol has shown a dual action pattern
491 depending on the dose; low concentrations increase expression of cell survival proteins,
492 whereas higher doses stimulate cell apoptosis or necrosis regardless of whether the cell
493 is healthy or pathological ^[91].

494 A recent study in humans showed that consumption of a phenolic-rich red grape
495 pomace drink produced an increase in the phenolic metabolites in plasma at different
496 times. Nevertheless, there was a high inter-individual variability in the absorption and

497 excretion of phenolic derivatives, which is a key point to be considered ^[93]. Gut
498 microbiota and phase II metabolic pathways play an important role in the biochemical
499 transformations of polyphenols and hence, more studies in humans are needed to
500 demonstrate the safety and therapeutic properties of dietary polyphenols. Furthermore, it
501 has been proposed that dietary intake of polyphenols derived from grape juice has a
502 beneficial effect on microbiota, which in turn can improve their bioavailability and
503 therefore contribute to enhancing human health benefits ^[95]. It is well known that the
504 polyphenol content in grape seed, skin or pomace extracts is higher than in grape juice,
505 so new procedures such as enzymatic treatment with exo-1,3- β -glucanase and
506 pectinases during juice obtention, could be used to improve the transfer of polyphenols
507 from the grape pomace ^[80, 81]. Other methods, such as microwave and ultrasound-
508 assisted extraction, pressurized solvent extraction, osmotic and membrane distillation,
509 heat and enzymatic digestion, nano and ultrafiltration, etc., have been also proved
510 effective to obtain extracts rich in polyphenols from grape and vine by-products ^[82-85].
511 Thus, it would be interesting to test the antileukemia properties of grape juice enriched
512 with polyphenol extracts ^[12, 86, 87].

513 The conclusion of this study is that grape polyphenols have a marked
514 antiproliferative effect on leukemia cells grown *in vitro*. However, further *in vivo*
515 studies are needed to improve the bioavailability of the compounds and to avoid the
516 toxicity produced at high concentrations. The interactions between the different
517 components present in natural or supplemented grape derived products could induce
518 antagonistic, synergistic, or additive effects that make it difficult to differentiate
519 between prevention and therapy for the treatment of leukemia. Once these important
520 points have been addressed and safety procedures standardized, clinical trials with

521 leukemia patients should be conducted in order to determine whether grape polyphenols
522 can be used as a preventive or therapeutic anti-leukemic natural product.

523

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530

531 Conflicts of Interest

532 The authors declare no conflict of interest.

533

534 **References**

- 535 1. Ignat, I.; Volf, I.; Popa, V. I., A critical review of methods for characterisation of
536 polyphenolic compounds in fruits and vegetables. *Food Chemistry* **2011**, *126* (4),
537 1821-1835.
- 538 2. Sobhani, M.; Farzaei, M. H.; Kiani, S.; Khodarahmi, R., Immunomodulatory: Anti-
539 inflammatory/antioxidant Effects of Polyphenols: A Comparative Review on the
540 Parental Compounds and Their Metabolites. *Food Reviews International* **2020**, 1-
541 53.
- 542 3. Martin, D., Los compuestos fenólicos: un acercamiento a su biosíntesis, síntesis y
543 actividad biológica. . *Rev Inv Agr y Amb* **2018**, *9* (1), 1-24.
- 544 4. Naumovski, N., Bioactive composition of plants and plant foods. 2015; pp 81-115.
- 545 5. Shi, J.; Yu, J.; Pohorly, J. E.; Kakuda, Y., Polyphenolics in grape seeds-
546 biochemistry and functionality. *J Med Food* **2003**, *6* (4), 291-299.
- 547 6. Perez-Jimenez, J.; Neveu, V.; Vos, F.; Scalbert, A., Identification of the 100 richest
548 dietary sources of polyphenols: an application of the Phenol-Explorer database. *Eur*
549 *J Clin Nutr* **2010**, *64 Suppl 3*, 112-120.
- 550 7. Perez-Jimenez, J.; Neveu, V.; Vos, F.; Scalbert, A., Systematic analysis of the
551 content of 502 polyphenols in 452 foods and beverages: an application of the
552 phenol-explorer database. *J Agric Food Chem* **2010**, *58* (8), 4959-4969.

- 553 8. Magrone, T.; Magrone, M.; Russo, M. A.; Jirillo, E., Recent Advances on the Anti-
554 Inflammatory and Antioxidant Properties of Red Grape Polyphenols: In Vitro and In
555 Vivo Studies. *Antioxidants* **2019**, *9* (1), 35-63.
- 556 9. Yahfoufi, N.; Alsadi, N.; Jambi, M.; Matar, C., The Immunomodulatory and Anti-
557 Inflammatory Role of Polyphenols. *Nutrients* **2018**, *10* (11), 1618-1631.
- 558 10. Rasouli, H.; Farzaei, M. H.; Khodarahmi, R., Polyphenols and their benefits: A
559 review. *Int. J. Food Prop* **2017**, *20*, 1700-1741.
- 560 11. Del Rio, D.; Rodriguez-Mateos, A.; Spencer, J. P. E.; Tognolini, M.; Borges, G.;
561 Crozier, A., Dietary (poly)phenolics in human health: structures, bioavailability, and
562 evidence of protective effects against chronic diseases. *Antioxidants & redox*
563 *signaling* **2013**, *18* (14), 1818-1892.
- 564 12. Gupta, M.; Dey, S.; Marbaniang, D.; Pal, P.; Ray, S.; Mazumder, B., Grape seed
565 extract: having a potential health benefits. *J. Food Sci Technol* **2020**, *57* (4), 1205-
566 1215.
- 567 13. Maia, M.; Ferreira, A. E. N.; Laureano, G.; Marques, A. P.; Torres, V. M.; Silva,
568 A. B.; Matos, A. R.; Cordeiro, C.; Figueiredo, A.; Sousa Silva, M., Vitis vinifera
569 'Pinot noir' leaves as a source of bioactive nutraceutical compounds. *Food &*
570 *Function* **2019**, *10* (7), 3822-3827.
- 571 14. Olmo-Cunillera, A.; Escobar-Avello, D.; Perez, A. J.; Marhuenda-Munoz, M.;
572 Lamuela-Raventos, R. M.; Vallverdu-Queralt, A., Is Eating Raisins Healthy?
573 *Nutrients* **2019**, *12* (1), 54-71.
- 574 15. Giovinazzo, G.; Grieco, F., Functional Properties of Grape and Wine Polyphenols.
575 *F. Plant Foods Hum Nutr* **2015**, *70*, 454-462.
- 576 16. Rauf, A.; Imran, M.; Butt, M. S.; Nadeem, M.; Peters, D. G.; Mubarak, M. S.,
577 Resveratrol as an anti-cancer agent: A review. *Crit Rev Food Sci Nutr* **2018**, *58* (9),
578 1428-1447.
- 579 17. Farzaei, M. H.; Bahramsoltani, R.; Rahimi, R., Phytochemicals as Adjunctive with
580 Conventional Anticancer Therapies. *Curr Pharm Des* **2016**, *22* (27), 4201-4218.
- 581 18. Silva, L. B. A. R.; Pinheiro-Castro, N.; Novaes, G. M.; Pascoal, G. d. F. L.; Ong,
582 T. P., Bioactive food compounds, epigenetics and chronic disease prevention: Focus
583 on early-life interventions with polyphenols. *Food Res Int* **2019**, *125* (108646), 1-
584 14.
- 585 19. Costa, C.; Tsatsakis, A.; Mamoulakis, C.; Teodoro, M.; Briguglio, G.; Caruso,
586 E.; Tsoukalas, D.; Margina, D.; Dardiotis, E.; Kouretas, D.; Fenga, C., Current
587 evidence on the effect of dietary polyphenols intake on chronic diseases. *Food*
588 *Chem Toxicol* **2017**, *110*, 286-299.
- 589 20. Curti, V.; Di Lorenzo, A.; Dacrema, M.; Xiao, J.; Nabavi, S. M.; Daglia, M.,
590 Review: In vitro polyphenol effects on apoptosis: An update of literature data.
591 *Seminars in Cancer Biology* **2017**, *46*, 119-131.
- 592 21. de Oliveira, M. R.; Nabavi, S. F.; Manayi, A.; Daglia, M.; Hajheydari, Z.;
593 Nabavi, S. M., Review: Resveratrol and the mitochondria: From triggering the
594 intrinsic apoptotic pathway to inducing mitochondrial biogenesis, a mechanistic
595 view. *BBA - General Subjects* **2016**, *1860*, 727-745.

- 596 22. Chojnacka, K.; Lewandowska, U., The Antiangiogenic Activity of Polyphenol-Rich
597 Extracts and Its Implication on Cancer Chemoprevention. *Food Reviews*
598 *International* **2020**, *36* (1), 77-103.
- 599 23. Huang, X.-t.; Li, X.; Xie, M.-l.; Huang, Z.; Huang, Y.-x.; Wu, G.-x.; Peng, Z.-r.;
600 Sun, Y.-n.; Ming, Q.-l.; Liu, Y.-x.; Chen, J.-p.; Xu, S.-n., Resveratrol: Review on
601 its discovery, anti-leukemia effects and pharmacokinetics. *Chemico-Biological*
602 *Interactions* **2019**, *306*, 29-38.
- 603 24. Grosso, G.; Godos, J.; Lamuela-Raventos, R.; Ray, S.; Micek, A.; Pajak, A.;
604 Sciacca, S.; D'Orazio, N.; Del Rio, D.; Galvano, F., A comprehensive meta-
605 analysis on dietary flavonoid and lignan intake and cancer risk: Level of evidence
606 and limitations. *Molecular Nutrition & Food Research* **2017**, *61* (4), 1-20.
- 607 25. Vanamala, J., Food systems approach to cancer prevention. *Critical Reviews in*
608 *Food Science and Nutrition* **2017**, *57* (12), 2573-2588.
- 609 26. Moga, M. A.; Dimienescu, O. G.; Arvatescu, C. A.; Mironescu, A.; Dracea, L.;
610 Ples, L., The Role of Natural Polyphenols in the Prevention and Treatment of
611 Cervical Cancer. An Overview. *Molecules* **2016**, *21* (8), 1-32.
- 612 27. NIH, N. C. I., Cancer Stats Fact; *Leukemia*. **2016**; pp 1-29.
- 613 28. Morceau, F.; Chateauvieux, S.; Orsini, M.; Trécul, A.; Dicato, M.; Diederich, M.,
614 Research review paper: Natural compounds and pharmaceuticals reprogram
615 leukemia cell differentiation pathways. *Biotechnology Advances* **2015**, *33* (1), 785-
616 797.
- 617 29. Harrison, T., *Oncología y hematología*. Principios de Medicina Interna: McGraw-
618 Hill Interamericana., **2006**.
- 619 30. Barrington-Trimis, J.; Cockburn, M.; Metayer, C.; Gauderman, W.; McKean-
620 Cowdin, R., Trends in Childhood Leukemia Incidence Over Two Decades from
621 1992-2013. *International journal of cancer* **2016**, *140* (5), 1-20.
- 622 31. Leon-Gonzalez, A.; José Jara-Palacios, M.; Abbas, M.; Heredia, F. J.; B Schini-
623 Kerth, V., Role of epigenetic regulation on the induction of apoptosis in Jurkat
624 leukemia cells by white grape pomace rich in phenolic compounds. *Food &*
625 *Function* **2017**, *8*, 4062-4069.
- 626 32. Espino, J.; Gonzalez-Gomez, D.; Moreno, D.; Fernández-León, M. F.; B
627 Rodríguez, A.; Pariente, J.; Delgado, J., Tempranillo-derived grape seed extract
628 induces apoptotic cell death and cell growth arrest in human promyelocytic
629 leukemia HL-60 cells. *Food & Function* **2013**, *4*, 1759-1766.
- 630 33. Athar, M.; Back, J. H.; Tang, X.; Kim, K. H.; Kopelovich, L.; Bickers, D. R.;
631 Kim, A. L., Review: Resveratrol: A review of preclinical studies for human cancer
632 prevention. *Toxicology and Applied Pharmacology* **2007**, *224*, 274-283.
- 633 34. Chen, J.; Tian, B.; Zhou, C.; Sun, J.; Lin, L.; Jin, S.; Liu, Q.; Fu, S.; Liu, L.;
634 Liu, H.; Zhang, Z.; Li, C.; Wei, H., A Novel Resveratrol-Arsenic Trioxide
635 Combination Treatment Synergistically Induces Apoptosis of Adriamycin-Selected
636 Drug-Resistant Leukemia K562 Cells. *Journal of Cancer* **2019**, *10* (22), 5483-5493.
- 637 35. Meng, J.; Liu, G. J.; Song, J. Y.; Chen, L.; Wang, A. H.; Gao, X. X.; Wang, Z. J.,
638 Preliminary results indicate resveratrol affects proliferation and apoptosis of

- 639 leukemia cells by regulating PTEN/PI3K/AKT pathway. *Eur Rev Med Pharmacol*
640 *Sci* **2019**, 23 (10), 4285-4292.
- 641 36. Siedlecka-Kroplewska, K.; Wozniak, M.; Kmiec, Z., The wine polyphenol
642 resveratrol modulates autophagy and induces apoptosis in MOLT-4 and HL-60
643 human leukemia cells. *J Physiol Pharmacol* **2019**, 70 (6), 825-838.
- 644 37. 40. Ferry-Dumazet, H.; Garnier, O.; Mamani-Matsuda, M.; Vercauteren, J.;
645 Belloc, F.; Billiard, C.; Dupouy, M.; Thiolat, D.; Kolb, J. P.; Marit, G.; Reiffers,
646 J.; Mossalayi, M. D., Resveratrol inhibits the growth and induces the apoptosis of
647 both normal and leukemic hematopoietic cells. *Carcinogenesis* **2002**, 23 (8), 1327-
648 1333.
- 649 38. Hutton, B.; Salanti, G.; Caldwell, D.; Chaimani, A.; Schmid, C.; Cameron, C.;
650 Ioannidis, J.; Straus, S.; Thorlund, K.; Jansen, J.; Mulrow, C.; Catalá-López, F.;
651 Getzsche, P.; Dickersin, K.; Boutron, I.; Altman, D.; Moher, D., The PRISMA
652 Extension Statement for Reporting of Systematic Reviews Incorporation Network
653 Meta-analyses of Health Care Interventions: Checklist and Explanation *Annals of*
654 *Internal Medicine*: **2015**; Vol. 162
- 655 39. Moher, D.; Shamseer L Fau - Clarke, M.; Clarke M Fau - Gherzi, D.; Gherzi D
656 Fau - Liberati, A.; Liberati A Fau - Petticrew, M.; Petticrew M Fau - Shekelle, P.;
657 Shekelle P Fau - Stewart, L. A.; Stewart, L. A., Preferred reporting items for
658 systematic review and meta-analysis protocols (PRISMA-P) 2015 statement. *Syst*
659 *Rev* **2015**, 4 (1), 1-9.
- 660 40. Shamseer, L.; Moher, D.; Clarke, M.; Gherzi, D.; Liberati, A.; Petticrew, M.;
661 Shekelle, P.; Stewart, L. A., Preferred reporting items for systematic review and
662 meta-analysis protocols (PRISMA-P) 2015: elaboration and explanation. *Bmj* **2015**,
663 349, 1-25.
- 664 41. Library, C. The Cochrane Library.
665 <https://www.cochranelibrary.com/es/search?cookiesEnabled>.
- 666 42. Elsevier EMBASE library. [https://www.elsevier.com/solutions/embase-biomedical-](https://www.elsevier.com/solutions/embase-biomedical-research)
667 [research](https://www.elsevier.com/solutions/embase-biomedical-research).
- 668 43. NIH MEDLINE Plus. U.S. National Library of Medicine. <https://medlineplus.gov>.
- 669 44. EBSCO The CINAHL database.
670 <https://www.ebscohost.com/nursing/products/cinahl-databases/cinahl-complete>
671 (accessed 1937).
- 672 45. Higgins, J.; Green, S., Cochrane Handbook for Systematic Reviews of Interventions
673 Version 5.1.0 Green, S., Ed. The Cochrane Collaboration: [www.cochrane-](http://www.cochrane-handbook.org)
674 [handbook.org](http://www.cochrane-handbook.org)., **2011**; Vol. 1, pp 1-639.
- 675 46. Veritas Health Innovation, M., Covidence systematic review software. Veritas
676 Health Innovation, Melbourne, Australia.: Veritas Health Innovation, Melbourne,
677 Australia., **2019**; Vol. 2001.
- 678 47. Centre, C. T. N. C. Review Manager (RevMan) [Computer program]. Version 5.3.
679 Copenhagen: The Nordic Cochrane Centre, The Cochrane Collaboration, **2014**.,
680 2014.
- 681 48. Socrates, A.; Pavlov D, S.; Clavero Ch, F., Riesgo relativo y Odds ratio ¿Qué son y
682 cómo se interpretan? *Rev Obstet Ginecol* **2010**, 5 (1), 51-54.

- 683 49. Edwardes, M.; Baltzan, M., The generalization of the odds ratio, risk ratio and risk
684 difference to $r \times k$ tables. *Statist Med* **2000**, *19*, 1901-1914.
- 685 50. Sierra, F., La sensibilidad y especificidad: entendiendo su origen y utilidad real. *Rev*
686 *Col Gastroenterol* **2003**, *18* (3), 180-182.
- 687 51. Billard, C.; Izard, J.-C.; Roman, V.; Kern, C.; Mathiot, C.; Mentz, F.; Kolb, J.-P.,
688 Comparative Antiproliferative and Apoptotic Effects of Resveratrol, ϵ -viniferin and
689 Vine-shots Derived Polyphenols (Vineatrols) on Chronic B Lymphocytic Leukemia
690 Cells and Normal Human Lymphocytes. *Leukemia & Lymphoma* **2002**, *43* (10),
691 1991-2002.
- 692 52. Can, G.; Cakir, Z.; Kartal, M.; Gunduz, U.; Baran, Y., Apoptotic effects of
693 resveratrol, a grape polyphenol, on imatinib-sensitive and resistant K562 chronic
694 myeloid leukemia cells. *Anticancer Res* **2012**, *32* (7), 2673-2678.
- 695 53. Chen, D.; Daniel, K. G.; Chen, M. S.; Kuhn, D. J.; Landis-Piwowar, K. R.; Dou,
696 Q. P., Dietary flavonoids as proteasome inhibitors and apoptosis inducers in human
697 leukemia cells. *Biochemical Pharmacology* **2005**, *69*, 1421-1432.
- 698 54. Gao, N.; Budhreja, A.; Cheng, S.; Yao, H.; Zhang, Z.; Shi, X., Induction of
699 apoptosis in human leukemia cells by grape seed extract occurs via activation of c-
700 Jun NH2-terminal kinase. *Clin Cancer Res* **2009**, *15* (1), 140-149.
- 701 55. Ghorbani, A.; Zand, H.; Jeddi-Tehrani, M.; Koohdani, F.; Shidfar, F.; Keshavarz,
702 S. A., PTEN over-expression by resveratrol in acute lymphoblastic leukemia cells
703 along with suppression of AKT/PKB and ERK1/2 in genotoxic stress. *Journal of*
704 *Natural Medicines* **2015**, *69* (4), 507-512.
- 705 56. Gokbulut, A. A.; Apohan, E.; Baran, Y., Resveratrol and quercetin-induced
706 apoptosis of human 232B4 chronic lymphocytic leukemia cells by activation of
707 caspase-3 and cell cycle arrest. *Hematology* **2013**, *18* (3), 144-150.
- 708 57. Hu, H.; Qin, Y.-m., Grape seed proanthocyanidin extract induced mitochondria-
709 associated apoptosis in human acute myeloid leukaemia 14.3D10 cells. *Chinese*
710 *Medical Journal* **2006**, *119* (5), 417-421.
- 711 58. Katsuzaki, H.; Hibasami, H.; Ohwaki, S.; Ishikawa, K.; Imai, K.; Date, K.;
712 Kimura, Y.; Komiya, T., Cyanidin 3-O-beta-D-glucoside isolated from skin of black
713 Glycine max and other anthocyanins isolated from skin of red grape induce
714 apoptosis in human lymphoid leukemia Molt 4B cells. *Oncology Reports* **2003**, *10*,
715 297-300.
- 716 59. Liao, H.-F.; Cheng, A.-J.; Huang, H.-T.; Shen, M.-L.; Hei, T. K.; Chen, Y.-J.,
717 Nuclear factor-kappaB as a switch in regulation of resveratrol-mediated apoptosis
718 and erythrocytic differentiation in human leukaemia cells. *Food Chemistry* **2012**,
719 *132* (4), 2094-2101.
- 720 60. Mertens-Talcott, S. U.; Percival, S. S.; Talcott, S. T., Extracts from red muscadine
721 and cabernet sauvignon wines induce cell death in MOLT-4 human leukemia cells.
722 *Food Chemistry* **2008**, *108* (3), 824-832.
- 723 61. Mertens-Talcott, S. U.; Talcott, S. T.; Percival, S. S., Low Concentrations of
724 Quercetin and Ellagic Acid Synergistically Influence Proliferation, Cytotoxicity and
725 Apoptosis in MOLT-4 Human Leukemia Cells. *The Journal of Nutrition* **2003**, *133*
726 (8), 2669-2674.

- 727 62. Sharif, T.; Auger, C.; Alhosin, M.; Ebel, C.; Achour, M.; Étienne-Selloum, N.;
728 Fuhrmann, G.; Bronner, C.; Schini-Kerth, V. B., Red wine polyphenols cause
729 growth inhibition and apoptosis in acute lymphoblastic leukaemia cells by inducing
730 a redox-sensitive up-regulation of p73 and down-regulation of UHRF1. *European*
731 *Journal of Cancer* **2010**, *46* (5), 983-994.
- 732 63. Siedlecka-Kroplewska, K.; Jozwik, A.; Kaszubowska, L.; Kowalczyk, A.;
733 Boguslawski, W., Pterostilbene induces cell cycle arrest and apoptosis in MOLT4
734 human leukemia cells. *Folia Histochemica et Cytobiologica* **2012**, *50* (4), 574-580.
- 735 64. Wang, M.; Wang, L.; Pan, X.-J.; Zhang, H., Monocytic differentiation of K562
736 cells induced by proanthocyanidins from grape seeds. *Archives of Pharmacol*
737 *Research* **2012**, *35* (1), 129-135.
- 738 65. Najaf Najafi, M.; Salehi, M.; Ghazanfarpour, M.; Hoseini, Z. S.; Khadem-
739 Rezaiyan, M., The association between green tea consumption and breast cancer
740 risk: A systematic review and meta-analysis. *Phytother Res* **2018**, *32* (10), 1855-
741 1864.
- 742 66. Gianfredi, V.; Vannini, S.; Moretti, M.; Villarini, M.; Bragazzi, N.; Izzotti, A.;
743 Nucci, D., Sulforaphane and epigallocatechin restore estrogen receptor expression
744 by modulating epigenetic events in the breast cancer cell line MDA-MB-231: a
745 systematic review and meta-analysis. *Journal of Nutrigenetics and Nutrigenomics*
746 **2017**, *10*, 126-135.
- 747 67. Feng, Y.; Zhou, J.; Jiang, Y., Resveratrol in lung cancer- a systematic review. *J*
748 *buon* **2016**, *21* (4), 950-953.
- 749 68. Chang, B.; Sang, L.; Wang, Y.; Tong, J.; Wang, B.-Y., Consumption of Tea and
750 Risk for Pancreatic Cancer: A Meta-Analysis of Published Epidemiological Studies.
751 *Nutrition and Cancer* **2014**, *66* (7), 1109-1123.
- 752 69. Imran, M.; Aslam, T.; Atif, M.; Shahbaz, M.; Qaisarani, T.; Salehi, B.;
753 Martorell, M.; Sharifi-Rad, J., Apigenin as an anticancer agent. *Phytotherapy*
754 *Research* **2020**, *1*, 1-17.
- 755 70. Salehi, B.; Venditti, A.; Sharifi-Rad, M.; Kręgiel, D.; Sharifi-Rad, J.; Durazzo,
756 A.; Lucarini, M.; Santini, A.; Souto, E. B.; Novellino, E.; Antolak, H.; Azzini,
757 E.; Setzer, W. N.; Martins, N., The Therapeutic Potential of Apigenin. *International*
758 *journal of molecular sciences* **2019**, *20* (6), 1305-1331.
- 759 71. Vrhovac Madunic, I.; Madunic, J.; Antunovic, M.; Paradzik, M.; Garaj-Vrhovac,
760 V.; Breljak, D.; Marijanovic, I.; Gajski, G., Apigenin, a dietary flavonoid, induces
761 apoptosis, DNA damage, and oxidative stress in human breast cancer MCF-7 and
762 MDA MB-231 cells. *Naunyn Schmiedebergs Arch Pharmacol* **2018**, *391* (5), 537-
763 550.
- 764 72. Lee, Y. H.; Chen, Y. Y.; Yeh, Y. L.; Wang, Y. J.; Chen, R. J., Stilbene
765 Compounds Inhibit Tumor Growth by the Induction of Cellular Senescence and the
766 Inhibition of Telomerase Activity. *Int J Mol Sci* **2019**, *20* (11), 1-22.
- 767 73. Ma, Z.; Zhang, X.; Xu, L.; Liu, D.; Di, S.; Li, W.; Zhang, J.; Zhang, H.; Li, X.;
768 Han, J.; Yan, X., Pterostilbene: Mechanisms of its action as oncostatic agent in cell
769 models and in vivo studies. *Pharmacological Research* **2019**, *145* (104265), 1-12.

- 770 74. Wen, W.; Lowe, G.; Roberts, C. M.; Finlay, J.; Han, E. S.; Glackin, C. A.;
771 Dellinger, T. H., Pterostilbene Suppresses Ovarian Cancer Growth via Induction of
772 Apoptosis and Blockade of Cell Cycle Progression Involving Inhibition of the
773 STAT3 Pathway. *Int J Mol Sci* **2018**, *19* (7), 1-12.
- 774 75. Gao, Y.; Tollefsbol, T. O., Combinational Proanthocyanidins and Resveratrol
775 Synergistically Inhibit Human Breast Cancer Cells and Impact Epigenetic(-
776)Mediating Machinery. *Int J Mol Sci* **2018**, *19* (8), 2204-2222.
- 777 76. Praud, D.; Parpinel, M.; Guercio, V.; Bosetti, C.; Serraino, D.; Facchini, G.;
778 Montella, M.; Vecchia, C.; Rossi, M., Proanthocyanidins and the risk of prostate
779 cancer in Italy. *Cancer Causes & Control* **2018**, *29*, 261-268.
- 780 77. Ravindranathan, P.; Pasham, D.; Balaji, U.; Cardenas, J.; Gu, J.; Toden, S.; Goel,
781 A., A combination of curcumin and oligomeric proanthocyanidins offer superior
782 anti-tumorigenic properties in colorectal cancer. *Scientific reports* **2018**, *8* (13869),
783 1-12.
- 784 78. Lionetti, V.; Tuana, B. S.; Casieri, V.; Parikh, M.; Pierce, G. N., Importance of
785 functional food compounds in cardioprotection through action on the epigenome.
786 *European Heart Journal* **2018**, *40* (7), 575-582.
- 787 79. Tzika, E.; Dreker, T.; Imhof, A., Epigenetics and Metabolism in Health and
788 Disease. *Frontiers in genetics* **2018**, *9*, 361-369.
- 789 80. Averilla, J. N.; Oh, J.; Wu, Z.; Liu, K. H.; Jang, C. H.; Kim, H. J.; Kim, J. S.,
790 Improved extraction of resveratrol and antioxidants from grape peel using heat and
791 enzymatic treatments. *J Sci Food Agric* **2019**, *99* (8), 4043-4053.
- 792 81. Lima Mdos, S.; da Conceicao Prudencio Dutra, M.; Toaldo, I. M.; Correa, L. C.;
793 Pereira, G. E.; de Oliveira, D.; Bordignon-Luiz, M. T.; Ninow, J. L., Phenolic
794 compounds, organic acids and antioxidant activity of grape juices produced in
795 industrial scale by different processes of maceration. *Food Chem* **2015**, *188*, 384-92.
- 796 82. Zwingelstein, M.; Draye, M.; Besombes, J. L.; Piot, C.; Chatel, G., Viticultural
797 wood waste as a source of polyphenols of interest: Opportunities and perspectives
798 through conventional and emerging extraction methods. *Waste Manag* **2020**, *102*,
799 782-794.
- 800 83. Nadar, S. S.; Rao, P.; Rathod, V. K., Enzyme assisted extraction of biomolecules as
801 an approach to novel extraction technology: A review. *Food Res Int* **2018**, *108*, 309-
802 330.
- 803 84. Cassano, A.; Conidi, C.; Ruby-Figueroa, R.; Castro-Munoz, R., Nanofiltration and
804 Tight Ultrafiltration Membranes for the Recovery of Polyphenols from Agro-Food
805 By-Products. *Int J Mol Sci* **2018**, *19* (2).
- 806 85. Al Bittar, S.; Perino-Issartier, S.; Dangles, O.; Chemat, F., An innovative grape
807 juice enriched in polyphenols by microwave-assisted extraction. *Food Chem* **2013**,
808 *141* (3), 3268-3272.
- 809 86. Li, F.-x.; Li, F.-h.; Yang, Y.-x.; Yin, R.; Ming, J., Comparison of phenolic profiles
810 and antioxidant activities in skins and pulps of eleven grape cultivars (*Vitis vinifera*
811 L.). *Journal of Integrative Agriculture* **2019**, *18* (5), 1148-1158.
- 812 87. Cheah, K. Y.; Howarth, G. S.; Bindon, K. A.; Kennedy, J. A.; Bastian, S. E. P.,
813 Low molecular weight procyanidins from grape seeds enhance the impact of 5-

- 814 Fluorouracil chemotherapy on Caco-2 human colon cancer cells. *PloS one* **2014**, 9
815 (6), 1-8.
- 816 88. Ramírez-Garza, S. L.; Laveriano-Santos, E. P.; Marhuenda-Muñoz, M.; Storniolo,
817 C. E.; Tresserra-Rimbau, A.; Vallverdú-Queralt, A.; Lamuela-Raventós, R. M.,
818 Health Effects of Resveratrol: Results from Human Intervention Trials. *Nutrients*
819 **2018**, 10 (12).
- 820 89. Francioso, A.; Mastromarino, P.; Masci, A.; d'Erme, M.; Mosca, L., Chemistry,
821 stability and bioavailability of resveratrol. *Med Chem* **2014**, 10 (3), 237-45.
- 822 90. Wenzel, E.; Somoza, V., Metabolism and bioavailability of trans-resveratrol. *Mol*
823 *Nutr Food Res* **2005**, 49 (5), 472-81.
- 824 91. San Hipólito-Luengo, Á.; Alcaide, A.; Ramos-González, M.; Cercas, E.; Vallejo,
825 S.; Romero, A.; Talero, E.; Sánchez-Ferrer, C. F.; Motilva, V.; Peiró, C., Dual
826 Effects of Resveratrol on Cell Death and Proliferation of Colon Cancer Cells. *Nutr*
827 *Cancer* **2017**, 69 (7), 1019-1027.
- 828 92. Wang, W.; Wang, Y. R.; Chen, J.; Chen, Y. J.; Wang, Z. X.; Geng, M.; Xu, D.
829 C.; Wang, Z. Y.; Li, J. H.; Xu, Z. D.; Pan, L. L.; Sun, J., Pterostilbene Attenuates
830 Experimental Atherosclerosis through Restoring Catalase-Mediated Redox Balance
831 in Vascular Smooth Muscle Cells. *J Agric Food Chem* **2019**, 67 (46), 12752-12760.
- 832 93. Castello, F.; Costabile, G.; Bresciani, L.; Tassotti, M.; Naviglio, D.; Luongo, D.;
833 Ciciola, P.; Vitale, M.; Vetrani, C.; Galaverna, G.; Brighenti, F.; Giacco, R.; Del
834 Rio, D.; Mena, P., Bioavailability and pharmacokinetic profile of grape pomace
835 phenolic compounds in humans. *Arch Biochem Biophys* **2018**, 646, 1-9.
- 836 94. Xu, W.; Yang, Y.; Xue, J. S.; Shi, J.; Lim, L.-T.; Forney, C.; Xu, G.; Bamba, S.
837 B., Effect of In Vitro Digestion on Water-in-Oil-in-Water Emulsions Containing
838 Anthocyanins from Grape Skin Powder. *Molecules* **2018**, 23 (11), 1-13.
- 839 95. Nash, V.; Ranadheera, C. S.; Georgousopoulou, E. N.; Mellor, D. D.;
840 Panagiotakos, D. B.; McKune, A. J.; Kellett, J.; Naumovski, N., The effects of
841 grape and red wine polyphenols on gut microbiota - A systematic review. *Food Res*
842 *Int* **2018**, 113, 277-287.
- 843

844 Table 1. Scientific articles included in the meta-analysis.

Authors	Year	O1	O2	O3
Billard et al. ^[51]	2002	X	X	
Can et al. ^[52]	2012	X	X	
Chen et al. ^[53]	2005	X	X	
Espino et al. ^[32]	2013			X
Ferry-Dumazet et al. ^[37]	2002	X	X	
Gao et al. ^[54]	2009	X	X	
Ghorbani et al. ^[55]	2015	X	X	
Gokbulut et al. ^[56]	2013	X	X	X
Hu et al. ^[57]	2006	X	X	
Katsuzaki et al. ^[58]	2003	X	X	
León-González et al. ^[31]	2017	X	X	
Liao et al. ^[59]	2012	X	X	X
Mertens-Talcott et al. ^[61]	2003	X	X	X
Mertens-Talcott et al. ^[60]	2008	X	X	X
Sharif et al. ^[62]	2010	X	X	
Siedlecka-Kroplewska et al. ^[63]	2012	X	X	X
Wang et al. ^[64]	2012	X	X	X

845

846 O1 (outcome 1): Effect of specific grape polyphenols on the proliferation of leukemia

847 cell lines. O2 (outcome 2): Effect of grape polyphenols on the proliferation of human

848 leukemia cells of different origin. O3 (outcome 3): Effect of grape polyphenols on cell

849 cycle arrest.

850

851 Table 2. Articles included in the meta-analysis showing the number of useful
 852 experiments for the data extraction, sorted by the type of leukemia cells and
 853 polyphenols.

854

Articles	T lymphocytes cell lines	Number of experiments		Articles	Grape polyphenols	Number of experiments
[58, 60, 61, 63]	MOLT-4	9		[31, 32, 54, 57, 60, 62, 64]	GPE	7
[31, 37, 53, 54]	JURKAT	8		[37, 51, 53, 55, 56, 59]	Resveratrol	6
	B lymphocytes cell lines			[53, 56, 61]	Quercetin	3
[37, 51]	WSU-CLL	4		[51]	Viniferin	3
[51]	ESKOL	3		[53]	Apigenin	1
[55]	NALM-6	1		[58]	Cyanidin glucoside	1
[54]	232B4	1		[58]	Delphinidin	1
	Early myeloid cell lines			[61]	Ellagic acid	1
[37, 52, 59, 64]	K562	4		[53]	Kaempferol	1
[32, 37, 54]	HL60	3		[58]	Malvidin	1
[57]	AML 14.3D10	1		[53]	Myricetin	1
[37]	KCL22	1		[58]	Petunidin	1
	Monocytic cell lines			[63]	Pterostilbene	1
[37, 54]	U937	2		[51]	Vineatrol	1
[37]	THP1	1				

855

856

857 Figure 1. Meta-analysis of the effect of grape polyphenols on leukemia cells'
858 proliferation, based on the PRISMA literature search guide ^[39].

859

860 Figure 2. Outcome 1's results of the effect of specific grape polyphenols on the
861 proliferation of human leukemia cells, grouped by type of polyphenol. Abbreviations:
862 Em, early myeloid cell lines; Mc, monocytic cell lines; LB, B lymphocytes cell lines;
863 LT, T lymphocytes cell lines.

864

865 Figure 3. Results of the effect of grape polyphenols on the proliferation of specific
866 human leukemia cells, grouped by cellular lineage. A. Effect of pure polyphenols. B.
867 Effect of GPE. Abbreviations: Em, early myeloid cell lines; Mc, monocytic cell lines;
868 LB, B lymphocytes cell lines; LT, T lymphocytes cell lines.

869

870 Figure 4. Effect of grape polyphenols on G₀ cell cycle arrest in leukemia cells.

871 Abbreviations: Em, early myeloid cell lines; LB, B lymphocytes cell lines; LT, T

872 Lymphocytes cell lines.