

DOES EXERCISE HAVE A NEUROPROTECTIVE FUNCTION IN MULTIPLE SCLEROSIS? A BRIEF OVERVIEW OF THE PHYSICAL TRAINING POTENTIAL EFFECTS ON CYTOKINES AND BRAIN-DERIVED NEUROTROPHIC FACTOR

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ABSTRACT

Although the advance in disease-modifying drugs has helped to stabilize the multiple sclerosis (MS) course increasing life-expectancy, physical deterioration still supervenes over time in most MS patients. In this context, physical exercise programs are considered a safe and well-tolerated tool to preserve functional independence in this population, which not only provides similar fitness improvements as usually observed in healthy general population, but it may also ameliorate some of the symptoms that this pathology entails (as fatigue, balance deficits, muscle weakness, etc.). Nowadays, the question is if exercise only aids to reverse physical deconditioning associated to the disease or it has the potential to have an impact on MS progression. In the present overview, the role of exercise as complementary therapy for modulating various physiopathological pathways related to MS disease such as inflammation and the neurotrophic support for neuronal survival was revised. Specifically, the exercise ability to modulate the immune system behaviour regulating the pro- and anti-inflammatory cytokine balance, as well as, to promote neuroprotective and neurorestorative mechanisms through the brain-derived neurotrophic factor stimulation was analysed.

Keywords: multiple sclerosis, exercise, cytokines, brain-derived neurotrophic factor, brain atrophy

¿PUEDE EL EJERCICIO FÍSICO EJERCER UNA FUNCIÓN NEUROPROTECTORA EN LA ESCLEROSIS MÚLTIPLE? UNA BREVE REVISIÓN DE LOS EFECTOS POTENCIALES DEL ENTRENAMIENTO FÍSICO SOBRE LAS CITOQUINAS Y EL FACTOR NEUROTROFICO DERIVADO DEL CEREBRO

RESUMEN

Aunque los avances farmacológicos han ayudado a estabilizar la evolución de la esclerosis múltiple (EM), aumentando notablemente la expectativa de vida, la mayoría de los pacientes con EM aún sufren un deterioro físico progresivo. En este contexto, los programas de ejercicio físico se consideran una herramienta segura que permite preservar la independencia funcional en esta población, proporcionando no solo las mejoras en la condición física que se observan en la población general, sino que también parecen eficaces para reducir la sintomatología asociada a la EM (como fatiga, déficit de equilibrio, debilidad muscular, etc.). Una de las preguntas actuales que existe en el ámbito científico es si el ejercicio físico sólo permite mejorar los síntomas y revertir el desacondicionamiento físico asociado con la EM o si también tiene el potencial de modular la progresión de la enfermedad. En la presente revisión se ofrece una perspectiva general del posible papel que puede jugar el ejercicio físico, como terapia complementaria en la regulación de varias

vías fisiopatológicas relacionadas con la EM, tales como la inflamación y el soporte neurotrófico para la supervivencia neuronal.

Específicamente, se analizó la capacidad que tiene el ejercicio físico para modular el comportamiento del sistema inmunitario a través de la regulación del equilibrio de citoquinas pro y anti-inflamatorias, así como para promover mecanismos neuroprotectores a través de la estimulación del factor neurotrófico derivado del cerebro.

Palabras clave: esclerosis múltiple, ejercicio, citoquinas, factor neurotrófico derivado del cerebro, atrofia cerebral

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INTRODUCTION

Multiple sclerosis (MS) is an immune-mediated chronic disease of the central nervous system (CNS) characterized by a breakdown of the blood-brain barrier (BBB), multifocal inflammation, demyelination, oligodendrocyte loss, reactive gliosis, and axonal degeneration (Baecher-Allan, Kaskow, & Weiner, 2018; Craig, Young, Ennis, Baker, & Boggild, 2003; Hohlfeld, 1997). MS neurodegeneration entails several physical symptoms such as balance disorders, muscular weakness, spasticity and fatigue (Gandhi, Laroni, & Weiner, 2010). Nowadays, MS is the most common neurological condition and it is among the most common causes of neurological disability in young adults in developed countries (Koch-Henriksen & Sorensen, 2010), with a much higher incidence (i.e. 3:1 ratio) among women (Orton et al., 2006). In Spain, its prevalence in the last 40 years has grown, affecting over 100 patients/100.000 people (Fernandez et al., 2012). Although the precise aetiology of the disease is unclear, the immune dysregulation seems to be prompted by an interaction between environmental and genetic factors (Hohlfeld, 1997; Koch-Henriksen & Sorensen, 2010). In individuals with a genetic predisposition to develop autoimmunity, MS is probably triggered by an environmental antigen (i.e. viral or bacterial infection), which auto-reactive T cells (Th1 and Th17), in a process identified as molecular mimicry, targets myelin antigens in the CNS (Garg & Smith, 2015; O'Connor, Bar-Or, & Hafler, 2001). The different pathophysiological mechanisms of the disease contribute to a brain volume loss, which seems to be accelerated in patients with MS [annual rate: 0.5–1.35% per year in patients with relapsing-remitting MS (RRMS)] compared to the healthy general population (annual rate: 0.1%–0.3%) (Calabrese et al., 2007; De Stefano et al., 2014; De Stefano et al., 2010; Hardmeier et al., 2005; Narayana et al., 2012; Rudick, Fisher, Lee, Simon, & Jacobs, 1999; Simon, 2006). Brain atrophy is a prominent feature of MS that advances inexorably throughout the disease course, with a rate that seems to be mostly independent of the MS subtype (De Stefano et al., 2014; De Stefano et al., 2010).

Although the improvement in disease-modifying drugs has helped to stabilize the disease progression increasing life-expectancy (Scalfari et al., 2013), physical deterioration still ensues over time in most MS patients (Filippini et al., 2013). In this context, rehabilitation programs become a key tool to preserve functional independence and quality of life (Dalgas, 2011). Among different complementary therapies, exercise, understood as a planned, structured and repetitive physical activity undertaken over a prolonged period (Caspersen, Powell, & Christenson, 1985), has shown to promote relevant, not only physical but also psychological and cognitive, improvements in MS patients (Aidar et al., 2018; Pilutti, Greenlee, Motl, Nickrent, & Petruzzello, 2013; Platta, Ensari, Motl, & Pilutti, 2016). Years ago, MS individuals were advised not

to perform any vigorous physical activity because it could worsen their neurological status (Giesser, 2015; Sutherland & Andersen, 2001). This is known as Uhthoff's phenomenon, in which some MS individuals show symptom exacerbations such as balance perturbations and increased spasticity when they become overheated during exercises (Dalgas, 2011; Giesser, 2015; Opara, Broła, Wylegala, & Wylegala, 2016). In addition, because patients generally experience symptoms of fatigue, they were also advised to avoid exercise in order to preserve their energy for activities of daily living. Both concerns usually made MS individuals be less active (Motl, McAuley, & Snook, 2005; Nortvedt, Riise, & Maeland, 2005), fostering physical deconditioning and potentially creating additional comorbidities such as obesity, osteoporosis or vascular diseases (Giesser, 2015). Nevertheless, it has already been shown that physical exercise does not increase the risk of relapse or adverse events in this population (Giesser, 2015; Pilutti, Platta, Motl, & Latimer-Cheung, 2014); conversely, exercise appears to be safe, well-tolerated and not only provides similar health and fitness improvements as usually observed in healthy individuals, but it also may ameliorate some of the symptoms that this pathology entails such as fatigue, balance deficits, muscle weakness, etc. (Cruickshank, Reyes, & Ziman, 2015; Giesser, 2015; Heine, van de Port, Rietberg, van Wegen, & Kwakkel, 2015; Latimer-Cheung et al., 2013; Motl & Pilutti, 2012; Motl & Sandroff, 2015).

An important question related to the impact of physical training in MS individuals' status is whether exercise only reverses physical deconditioning associated to the disease or it has a neuroprotective effect, which could attenuate the disease progression (Dalgas & Stenager, 2012). The data from the last decade reinforce the second approach, suggesting that physical activity could modulate to some extent various physiopathological pathways related to MS disease, like inflammation and the neurotrophic support for neuronal survival and regeneration.

CNS inflammation in MS: the potential effect of exercise on pro- and anti-inflammatory cytokines

According to McDonald's criteria (Thompson et al., 2018), most MS patients suffer the RRMS form of the disease, which is initially characterized by relapses of focal neurological deficits followed by a variable degree of recovery (Compston & Coles, 2008). During the early stage of the RRMS, CNS inflammation seems to be the primary cause of damage (Loma & Heyman, 2011). This phase is probably driven by a systemic immune response, in which there is a myelin-specific autoreactive lymphocyte infiltration of the BBB that leads to the formation of inflammatory demyelinating lesions (Cicarelli et al., 2014; Olsson, 1995). Cytokines are proteins that regulate the immune system

function (Alexander, 2002) playing an important role in the pathogenic events that characterize MS progression (Ozenci, Kouwenhoven, & Link, 2002). Commonly, in healthy individuals, due to a cross-inhibitory effect, there is a homeostatic balance between pro-and anti-inflammatory cytokines. In presence of pathogens this balance changes, resulting in an increase of pro-inflammatory cytokines [i.e. interleukin (IL)-1, IL-6, IL-12, IL-17, IL-23 tumor necrosis factor (TNF)- α , interferon (INF)- γ] and a decrease of the anti-inflammatory ones (i.e. IL-4, IL-10, IL-13) which induces, through different pathways, a local inflammatory and immunological response (i.e. immune cell differentiation, growth, proliferation and activation) (Ozenci et al., 2002). However, as the immune cells are sensitized to the myelin antigen in MS, the pro-inflammatory imbalance is self-sustained over time which induces the cascade of inflammatory and demyelination processes observed in the illness pathogenesis (Pilli, Zou, Tea, Dale, & Brilot, 2017): 1) Pro-inflammatory cytokines IL-12 and IL-23 participate in the peripheral immune activation (differentiation of CD4+ T cell into Th1 or Th17 phenotypes) regulating the interaction between T lymphocytes and the antigen presenting cells such as B cells, dendritic cells, microglia and macrophages (Loma & Heyman, 2011); 2) Cytokines also facilitate the migration of those activated T cells into the CNS disrupting the BBB through TNF- α , INF γ , IL-1 β , IL-17 and IL-22 (Kebir et al., 2007; Minagar & Alexander, 2003); 3) Once activated immune cells have crossed the BBB, they release several pro-inflammatory cytokines stimulating microglia, macrophages, astrocytes, and recruit B cells, which, in turn, damage myelin, oligodendrocytes and axons (Rivera-Quinones et al., 1998; Zamvil & Steinman, 2003). It must be noted that the release of pro-inflammatory cytokines such as TNF- α and INF γ promotes inflammation, also suppressing the differentiation of T-regulatory cells (Th2), which release anti-inflammatory cytokines (Minty et al., 1993; Zhu & Paul, 2008).

Due to the role of cytokines in MS pathogenesis, the balance regulation of the pro- and anti-inflammatory cytokines is a major issue, not only for disease-modifying drugs therapies (Motl et al., 2017) but for any complementary treatment trying to slow-down the disease progression. In this sense, as regular exercise has shown to chronically reduce the basal levels of inflammatory markers (Petersen & Pedersen, 2005), it may be expected that exercise could be used as an anti-inflammatory and complementary therapy in MS patients (Dalgas & Stenager, 2012). Physical activity seems to reduce Th1 cell production, but not Th2 cells production (Gleeson, 2007; Sharif et al., 2018; Steensberg et al., 2001), promoting a long-term immune regulation characterized by a post-exercise transient anti-inflammatory state (Gleeson et al., 2011). During and after exercise, IL-6 is released from muscles, which, in this case, acts as a myokine, inducing an anti-inflammatory response (Brandt &

Pedersen, 2010; Sharif et al., 2018). Commonly, IL-6 works together with TNF- α as a pro-inflammatory cytokine against a pathogen, promoting the proliferation and activation of Th1 cells (Sharif et al., 2018). Nevertheless, after exercise, IL-6 is released from muscles without TNF- α secretion, working as a myokine. In this condition, IL-6 promotes the secretion of IL-10 and the reduction of TNF- α , reducing inflammation and inhibiting Th1-cells (de Vries, 1995). Conversely, IL-6 from monocytes, which acts as a pro-inflammatory cytokine, as well as IL-2 from Th1 lymphocytes, are inhibited during and after exercise, contributing to the inflammatory reduction (Lancaster et al., 2004). The decrease of Th1 following exercise is also attributed to the increase in hormonal levels of cortisol and adrenaline in response to physical activity, which inhibits the production of IL-12 from antigen presenting cells (Elenkov & Chrousos, 1999; Pedersen & Hoffman-Goetz, 2000) and reduces TNF- α produced by Th1 cells (Bergmann et al., 1999). Although changes in circulating lymphocytes normally return to baseline levels in no more than 24 h (Gleeson, 2007), systematic changes in IL-6, cortisol and adrenaline caused by regular bouts of exercise could offer protection against auto-immune disease (Gleeson, 2007; Lancaster et al., 2004).

While the effectiveness of regular exercise to modulate the cytokine profile and the progression of inflammatory-related diseases has been proven (Bilski et al., 2016; Leal, Lopes, & Batista, 2018; Pearson, Mungovan, & Smart, 2018; Pedersen, 2017), experimental studies on MS have shown limited and contradictory results. In a recent systematic review of the experimental studies conducted by Negaresh et al. (2018), no systematic cytokine changes in serum associated to regular exercise were found, which, according to the authors, would not support the idea that the cytokine modifications caused by exercise are a regulator of the disease progression. Nevertheless, the real effect of exercise on cytokine in MS must still be elucidated because, as the aforementioned authors pointed out, the few experimental studies present several methodological limitations, which hinder the generalization of the results: small samples, only female population, different MS types in the same experimental group, cytokines monitored using a variety of methods and taken from different body fluids, etc. For example, the fact that cytokine levels in plasma and serum did not change towards a more anti-inflammatory profile can be attributed to a lower BBB permeability caused by exercise (Mokhtarzade et al., 2018) which would limit the immune cell infiltration into the CNS.

Among those limitations, from the point of view of physical exercise prescription, one of the most important ones that could explain the current disparity of cytokine findings was the high heterogeneity observed in the exercise programs. Acute and chronic cytokine modifications caused by exercise depend on several parameters as, for example, the type of exercise (i.e.

aerobic training, resistance training, combined training, etc.), the intensity and volume of training, program duration, participant training status, age, type of the disorder, etc. (Gleeson, 2007). For instance, focusing on the secretion of IL-6 induced by exercise, it has been observed that the higher duration, intensity and greater number of muscles involved in the exercise are, the greater the release of this cytokine after exercise is (Gokhale, Chandrashekara, & Vasanthakumar, 2007; Pedersen & Febbraio, 2005). Probably, longer and more intense training implies higher energy demands and consequently, a greater carbohydrate depletion associated to a higher acute IL-6 production from muscles (Keller et al., 2001) which, in turn, could chronically modify pro- and anti-inflammatory cytokine profile. This rationale could explain why some studies found significant changes in the cytokine levels after training in RRMS individuals (Deckx et al., 2016; Golzari, Shabkhiz, Soudi, Kordi, & Hashemi, 2010; Kierkegaard et al., 2016; Mokhtarzade, Ranjbar, Majdinasab, Patel, & Molanouri Shamsi, 2017). For instance, Kierkegaard et al. (2016) found a significant reduction in the basal level of several pro-inflammatory cytokines in serum in RRMS after a high-intensity resistance training performed at 80% of one repetition maximum test involving several muscle groups (8 exercises) during three months (24 sessions). Golzari et al. (2010) observed a significant reduction in plasma INF γ and IL-17 after a combined endurance and resistance training of 3 sessions per week for 2 months (24 sessions). Similarly, Deckx et al. (2016) found a significant reduction in plasma TNF- α when faced with lipopolysaccharide and IFN- γ stimulation after a combined endurance and resistance training of 5 sessions per 2 weeks for 3 months (30 sessions). Overall, these findings suggest that the higher the duration, intensity and energy demand are, the greater the cytokine activity is. However, as individuals with higher fitness level show a reduced response to exercise (Gokhale et al., 2007; Pedersen & Febbraio, 2005), a continuous and individualized modulation of training parameters seems to be required to elicit cytokine adaptations along time. In spite of these potential implications, nowadays, the small number of experimental studies carried out on MS, the heterogeneity of the experimental design and the inconsistent reporting of their training parameters hinder the dose-response characterization of the exercise effect on cytokines in this population (Negaresh et al., 2018).

Additionally, another limitation related to the exercise program design was whether the training protocols were enough to induce weight loss (Mokhtarzade et al., 2017; Negaresh et al., 2018), which is an important tool to regulate the secretion of the cytokines by the adipose tissue, i.e., adipokines (Gleeson et al., 2011). An important adipokine related to the MS pathology is leptin, which promotes the secretion of pro-inflammatory cytokines TNF- α , IL-2, and IL-6 and the activation of Th1 cells (Matarese, Procaccini, & De Rosa, 2008).

In this sense, Mokhtarzade et al. (2017) observed that weight loss in RRMS induced by an exercise program was significantly related with a reduction in leptin together with TNF- α , which, in turn, were related with greater fatigue improvements. These authors also observed an increase in the adiponectin hormone which increases the production of the anti-inflammatory cytokine IL-10 (Kraszula et al., 2012). Although these preliminary findings suggest that exercise programs on MS should also focus on weight loss in order to maximize their effects on the cytokine and adipokine profile, further research is necessary to clarify the role of weight loss after training on the immune system in MS.

CNS integrity in MS: the potential effect of exercise on brain-derived neurotrophic factor

Although white matter infiltration by immune cells and subsequent demyelination are commonly considered the most recognizable signs of MS, demyelination of the cortical gray matter, axonal deterioration and CNS atrophy have been detected in MS patients even in early stages of the disease (Baecher-Allan et al., 2018; Bevan et al., 2018; Kutzelnigg et al., 2005; Lucchinetti et al., 2011; Siffirin, Vogt, Radbruch, Nitsch, & Zipp, 2010; Trapp & Nave, 2008). In MS patients, brain atrophy, mainly assessed through magnetic resonance imaging techniques, progresses at a faster rate than in healthy individuals (Vollmer et al., 2015). Importantly, neurodegenerative rather than demyelination processes seem to be more related with the degree of disability in MS (Lemus, Warrington, & Rodriguez, 2018; Wujek et al., 2002). In rodent models, no motor impairment has been observed in presence of demyelination if the axonal integrity was preserved (Rivera-Quinones et al., 1998), but long-lasting neurological disability was found in presence of axonal loss (Wujek et al., 2002). These findings have also been observed in MS patients, in whom the cortical and gray matter atrophy was the best predictor of the disease progression and motor disability (Fisniku et al., 2008) as well as of the degree of cognitive impairment (Sacco et al., 2015). In this sense, it has been suggested that the evolution from RRMS to a progressive form of the disease materializes when the CNS is not able to compensate the axonal injuries (Ksiazek-Winiarek, Szpakowski, & Glabinski, 2015; Nave & Trapp, 2008). This explains why there is a higher rate of axonal loss and brain atrophy in later stages of the disease (Simon, 2006). All these reasons justify that therapeutic interventions aim at not only reducing relapse events, inflammation and demyelination but also at enhancing neuroprotection and neurorestoration of the CNS (Luhder, Gold, Flugel, & Linker, 2013).

A therapeutic target for preserving CNS integrity are the neurotrophins, which are proteins involved in neural regeneration, preservation and

remyelination processes (Azoulay, Vachapova, Shihman, Miler, & Karni, 2005; Begni, Riva, & Cattaneo, 2017; Ceni, Unsain, Zeinieh, & Barker, 2014; Ebadi et al., 1997; Huang & Reichardt, 2001; Kelamangalath & Smith, 2013; Linker, Gold, & Luhder, 2009; Vacaras, Major, & Buzoianu, 2017). One of the most important neurotrophins is the brain-derived neurotrophic factor (BDNF) which, in healthy subjects, has been associated with age-related volume reduction in white matter (Driscoll et al., 2012) and brain volume, mostly the hippocampus and the amygdala (Erickson, Prakash, et al., 2010; Manna et al., 2015). BDNF is also involved in the pathogenesis of several neurological diseases such as Huntington's disease (Ferrer, Goutan, Marin, Rey, & Ribalta, 2000; Zuccato & Cattaneo, 2007), Alzheimer's disease (Ye, Tai, & Zhang, 2012) and Parkinson's disease (Howells et al., 2000). Mature BDNF functions are related to several neuroplasticity mechanisms as neuronal protection and survival, axonal and dendritic remodeling, neuronal differentiation, synaptogenesis in axon terminals and synaptic transmission efficacy (Alsina, Vu, & Cohen-Cory, 2001; Barde, 1994; Knaepen, Goekint, Heyman, & Meeusen, 2010). The major source of circulating BDNF (70–80%) is the brain, mainly the cortex and hippocampus (Rasmussen et al., 2009). Commonly, serum and plasma levels of BDNF, which are highly correlated to BDNF in the CNS (Pan, Banks, Fasold, Bluth, & Kastin, 1998), are reduced in MS patients compared to healthy controls (Azoulay et al., 2005; Tongiorgi et al., 2012). However, during periods of higher inflammatory activity, especially after MS relapses (Frota et al., 2009), BDNF levels rise (Weinstock-Guttman et al., 2007) being observed not only in plasma but also in immune and CNS cells of demyelinating lesions (Stadelmann et al., 2002). This transitory increase of BDNF associated to active stages of the disease is produced not only by neurons but also by immune cells (T lymphocyte, microglia and astrocytes) (Kerschensteiner et al., 1999), which has been interpreted as a neuroprotective mechanism in response to CNS injuries. The neuroprotective role of the BDNF in MS would also be supported by correlational results, which have showed that the greater the BDNF release is, the higher the white matter volume in MS patients is (Weinstock-Guttman et al., 2007). Thus, a lower BDNF secreted by neuron and immune cells linked to the disease progression would indicate an exhaustion of the CNS restoration ability (Azoulay, Urshansky, & Karni, 2008; Azoulay et al., 2005).

It is currently well recognized that exercise elicits molecular and cellular processes that promote neural plasticity in aging and disease (White & Castellano, 2008). Interestingly, among the potential pathways for CNS preservation, BDNF seems to be the most susceptible neurotrophin to be modulated by exercise (Knaepen et al., 2010). Commonly, BDNF circulating levels from several brain areas such as the hippocampus, amygdala, prefrontal and motor cortex, cerebellum, etc. (Fang et al., 2013; S. Gustafsson, Liang, &

Hilke, 2011; Koo et al., 2013; Liu et al., 2009; Marais, Stein, & Daniels, 2009; Neeper, Gomez-Pinilla, Choi, & Cotman, 1996) rise after a single bout of exercise (Ferris, Williams, & Shen, 2007; G. Gustafsson et al., 2009; Rojas Vega et al., 2006) and they generally return back to baseline values in less than 1 h (Knaepen et al., 2010). As various meta-analyses have confirmed (Dinoff et al., 2016; Szuhany, Bugatti, & Otto, 2015), exercise training programs chronically increase resting concentrations of BDNF in peripheral blood in healthy adults and, to higher extent, in individuals with neurological disorders as Parkinson's disease (Hirsch, van Wegen, Newman, & Heyn, 2018). It must be noted that aerobic rather than resistance training programs seem to be more effective to increase resting BDNF concentrations in peripheral blood (Dinoff et al., 2016), but probably BDNF modulation depends on the type of exercise, number of muscles involved, intensity and duration of the exercise bouts as wells as the participants' physical status. BDNF changes caused by regular exercise could explain why higher fitness levels and physical activity are associated with greater brain volume. Cross-sectional studies (Colcombe et al., 2003; Erickson et al., 2009; Floel et al., 2010; Verstynen et al., 2012) but more importantly longitudinal works (Erickson, Raji, et al., 2010) as well as randomized control trials (Colcombe et al., 2006; Erickson et al., 2011; Ruscheweyh et al., 2011) have shown how physical fitness and exercise are linked with brain volume and gray matter preservation in older adults (Erickson, Leckie, & Weinstein, 2014). Interestingly, as Erickson et al. (2011) showed, higher serum BDNF changes caused by an aerobic training program were correlated with changes in the hippocampal volume, which would confirm the neuroprotective role of BDNF as a physiological mediator of exercise effects on CNS preservation. It is important to note that a similar association has been found between physical fitness and white matter volumes, not only in older adults (Sexton et al., 2016), but also in children (Chaddock-Heyman et al., 2014).

Based on these findings, recent works have tried to elucidate if exercise can work in a similar way in MS patients, helping to slow-down demyelination and axonal deterioration caused by the disease. To the best of the authors' knowledge, the first study which observed the link between exercise and MS progression was carried out by Prakash, Snook, Motl, and Kramer (2010). In this study, a significant relationship between aerobic fitness and both, gray matter volume and white matter integrity in RRMS patients was found. Subsequent cross-sectional studies have confirmed these results finding significant associations between the physical activity level and the volume of some brain areas (Klaren et al., 2015; Schwartz et al., 2016). These results, although promising, can be biased by confounding variables, especially the stage of the disease in which each patient is. Thus, longitudinal studies and randomized trials are needed to reduce the influence of confounding

parameters. To the best of the authors' knowledge, only three intervention studies have analysed the effect of exercise programs on brain volume parameters in MS. The first one was a case study (Leavitt et al., 2014), in which an RRMS patient, who was involved in a 3-month aerobic training program, showed an important increase of the hippocampal volume (+16.5%), while another RRMS patient, who underwent a stretching program, did not show any noticeable change (+2.8%). Later, a very recent pilot study found a lower but non-significant ($p < 0.1$) brain atrophy in a group of RRMS patients who carried out a 6-month resistance training program compared with a control group (Kjohede et al., 2018). Finally, the first randomized control trial analysing exercise and brain atrophy found that RRMS patients involved in a community-located running training program presented an increase in brain volume compared with a control group; however, these changes were only significant in the pallidum (Feys et al., 2017).

In spite of these promising results suggesting that exercise is not only helpful to reverse physical deconditioning associated to the MS disease but that it also has neuroprotective effects, from the authors' point of view, it is too early to assert that exercise is able to reduce the brain atrophy caused by MS progression. It is even more difficult to perform any recommendations about which type of training would be the most appropriate to achieve this goal. Many more experimental studies with a larger sample size and longer training duration are needed to clarify all these questions. Until that happens, sport science professionals, exercise physiologists and physiotherapists should base their physical exercise interventions on the general recommendations and precautions developed for this population (Dalgas, Stenager, & Ingemann-Hansen, 2008; Halabchi, Alizadeh, Sahraian, & Abolhasani, 2017), which ensure that exercise is well-tolerated and focuses on the improvement of the most prominent symptoms of each patient.

CONCLUSIONS

Overall, the results from the last decade presented in this overview suggest that physical exercise might be a complementary but powerful therapy to regulate the immune system behaviour in MS through the modulation of pro- and anti-inflammatory cytokine balance, as well as to promote neuroprotective and neurorestorative mechanisms through BDNF stimulation. Therefore, although exercise does not make the front pages of the main scientific journals as frequently as drug research does, these findings on exercise reinforce the idea that more randomized controlled trials, supervised by a multidisciplinary group (neurologists, practitioners, sport science professionals, etc.), should be carried out to confirm if exercise can play a neuroprotective role capable to

slow-down MS course, and to quantify their dose-response association in both, the relapse-remitting and the progressive forms of the disease.

REFERENCES

- Aidar, F. J., Carneiro, A. L., Costa Moreira, O., Patrocínio de Oliveira, C. E., Garrido, N. D., Machado Reis, V., & Gama de Matos, D. (2018). Effects of resistance training on the physical condition of people with multiple sclerosis. *The Journal of Sports Medicine and Physical Fitness*, *58*(7-8), 1127-1134. doi: 10.23736/S0022-4707.17.07621-6.
- Alexander, W. S. (2002). Suppressors of cytokine signalling (SOCS) in the immune system. *Nature Reviews. Immunology*, *2*(6), 410-416. doi: 10.1038/nri818.
- Alsina, B., Vu, T., & Cohen-Cory, S. (2001). Visualizing synapse formation in arborizing optic axons in vivo: dynamics and modulation by BDNF. *Nature Neuroscience*, *4*(11), 1093-1101. doi: 10.1038/nn735.
- Azoulay, D., Urshansky, N., & Karni, A. (2008). Low and dysregulated BDNF secretion from immune cells of MS patients is related to reduced neuroprotection. *Journal of Neuroimmunology*, *195*(1-2), 186-193. doi: 10.1016/j.jneuroim.2008.01.010.
- Azoulay, D., Vachapova, V., Shihman, B., Miler, A., & Karni, A. (2005). Lower brain-derived neurotrophic factor in serum of relapsing remitting MS: reversal by glatiramer acetate. *Journal of Neuroimmunology*, *167*(1-2), 215-218. doi: 10.1016/j.jneuroim.2005.07.001.
- Baecher-Allan, C., Kaskow, B. J., & Weiner, H. L. (2018). Multiple Sclerosis: Mechanisms and Immunotherapy. *Neuron*, *97*(4), 742-768. doi: 10.1016/j.neuron.2018.01.021.
- Barde, Y. A. (1994). Neurotrophins: a family of proteins supporting the survival of neurons. *Progress in Clinical and Biological Research*, *390*, 45-56.
- Begni, V., Riva, M. A., & Cattaneo, A. (2017). Cellular and molecular mechanisms of the brain-derived neurotrophic factor in physiological and pathological conditions. *Clinical Science*, *131*(2), 123-138. doi: 10.1042/CS20160009.
- Bergmann, M., Gornikiewicz, A., Sautner, T., Waldmann, E., Weber, T., Mittlbock, M., & Fugger, R. (1999). Attenuation of catecholamine-induced immunosuppression in whole blood from patients with sepsis. *Shock*, *12*(6), 421-427.
- Bevan, R. J., Evans, R., Griffiths, L., Watkins, L. M., Rees, M. I., Magliozzi, R., & Howell, O. W. (2018). Meningeal Inflammation and Cortical Demyelination in Acute Multiple Sclerosis. *Annals of Neurology*. doi: 10.1002/ana.25365.
- Bilski, J., Mazur-Bialy, A., Brzozowski, B., Magierowski, M., Zahradnik-Bilska, J., Wojcik, D., & Brzozowski, T. (2016). Can exercise affect the course of

- inflammatory bowel disease? Experimental and clinical evidence. *Pharmacological Reports*, 68(4), 827-836.
doi: 10.1016/j.pharep.2016.04.009.
- Brandt, C., & Pedersen, B. K. (2010). The role of exercise-induced myokines in muscle homeostasis and the defense against chronic diseases. *Journal of Biomedicine & Biotechnology*, 2010, 520258. doi: 10.1155/2010/520258.
- Calabrese, M., Atzori, M., Bernardi, V., Morra, A., Romualdi, C., Rinaldi, L., & Gallo, P. (2007). Cortical atrophy is relevant in multiple sclerosis at clinical onset. *Journal of Neurology*, 254(9), 1212-1220. doi: 10.1007/s00415-006-0503-6.
- Caspersen, C. J., Powell, K. E., & Christenson, G. M. (1985). Physical activity, exercise, and physical fitness: definitions and distinctions for health-related research. *Public Health Reports*, 100(2), 126-131.
- Ceni, C., Unsain, N., Zeinieh, M. P., & Barker, P. A. (2014). Neurotrophins in the regulation of cellular survival and death. *Handbook of Experimental Pharmacology*, 220, 193-221. doi: 10.1007/978-3-642-45106-5_8.
- Ciccarelli, O., Barkhof, F., Bodini, B., De Stefano, N., Golay, X., Nicolay, K., & Miller, D. H. (2014). Pathogenesis of multiple sclerosis: insights from molecular and metabolic imaging. *Lancet Neurology*, 13(8), 807-822. doi: 10.1016/S1474-4422(14)70101-2.
- Colcombe, S. J., Erickson, K. I., Raz, N., Webb, A. G., Cohen, N. J., McAuley, E., & Kramer, A. F. (2003). Aerobic fitness reduces brain tissue loss in aging humans. *The Journals of Gerontology. Series A, Biological Sciences and Medical Sciences*, 58(2), 176-180.
- Colcombe, S. J., Erickson, K. I., Scalf, P. E., Kim, J. S., Prakash, R., McAuley, E., & Kramer, A. F. (2006). Aerobic exercise training increases brain volume in aging humans. *The Journals of Gerontology. Series A, Biological Sciences and Medical Sciences*, 61(11), 1166-1170.
- Compston, A., & Coles, A. (2008). Multiple sclerosis. *Lancet*, 372(9648), 1502-1517. doi: 10.1016/S0140-6736(08)61620-7.
- Craig, J., Young, C. A., Ennis, M., Baker, G., & Boggild, M. (2003). A randomised controlled trial comparing rehabilitation against standard therapy in multiple sclerosis patients receiving intravenous steroid treatment. *Journal of Neurology, Neurosurgery, and Psychiatry*, 74(9), 1225-1230.
- Cruikshank, T. M., Reyes, A. R., & Ziman, M. R. (2015). A systematic review and meta-analysis of strength training in individuals with multiple sclerosis or Parkinson disease. *Medicine*, 94(4), e411.
doi: 10.1097/MD.0000000000000411.
- Chaddock-Heyman, L., Erickson, K. I., Holtrop, J. L., Voss, M. W., Pontifex, M. B., Raine, L. B., & Kramer, A. F. (2014). Aerobic fitness is associated with greater white matter integrity in children. *Frontiers in Human Neuroscience*, 8, 584.
doi: 10.3389/fnhum.2014.00584.

- Dalgas, U. (2011). Rehabilitation and multiple sclerosis: hot topics in the preservation of physical functioning. *Journal of the Neurological Sciences*, *311 Suppl 1*, S43-47. doi: 10.1016/S0022-510X(11)70008-9.
- Dalgas, U., & Stenager, E. (2012). Exercise and disease progression in multiple sclerosis: can exercise slow down the progression of multiple sclerosis? *Therapeutic Advances in Neurological Disorders*, *5*(2), 81-95. doi: 10.1177/1756285611430719.
- Dalgas, U., Stenager, E., & Ingemann-Hansen, T. (2008). Multiple sclerosis and physical exercise: recommendations for the application of resistance-, endurance- and combined training. *Multiple Sclerosis*, *14*(1), 35-53. doi: 10.1177/1352458507079445.
- De Stefano, N., Airas, L., Grigoriadis, N., Mattle, H. P., O'Riordan, J., Oreja-Guevara, C., & Kieseier, B. C. (2014). Clinical relevance of brain volume measures in multiple sclerosis. *CNS Drugs*, *28*(2), 147-156. doi: 10.1007/s40263-014-0140-z.
- De Stefano, N., Giorgio, A., Battaglini, M., Rovaris, M., Sormani, M. P., Barkhof, F., & Filippi, M. (2010). Assessing brain atrophy rates in a large population of untreated multiple sclerosis subtypes. *Neurology*, *74*(23), 1868-1876. doi: 10.1212/WNL.0b013e3181e24136.
- de Vries, J. E. (1995). Immunosuppressive and anti-inflammatory properties of interleukin 10. *Annals of Medicine*, *27*(5), 537-541. doi: 10.3109/07853899509002465.
- Deckx, N., Wens, I., Nuyts, A. H., Hens, N., De Winter, B. Y., Koppen, G., & Cools, N. (2016). 12 Weeks of Combined Endurance and Resistance Training Reduces Innate Markers of Inflammation in a Randomized Controlled Clinical Trial in Patients with Multiple Sclerosis. *Mediators of Inflammation*, *2016*, 6789276. doi: 10.1155/2016/6789276.
- Dinoff, A., Herrmann, N., Swardfager, W., Liu, C. S., Sherman, C., Chan, S., & Lanctot, K. L. (2016). The Effect of Exercise Training on Resting Concentrations of Peripheral Brain-Derived Neurotrophic Factor (BDNF): A Meta-Analysis. *PloS One*, *11*(9), e0163037. doi: 10.1371/journal.pone.0163037.
- Driscoll, I., Martin, B., An, Y., Maudsley, S., Ferrucci, L., Mattson, M. P., & Resnick, S. M. (2012). Plasma BDNF is associated with age-related white matter atrophy but not with cognitive function in older, non-demented adults. *PloS One*, *7*(4), e35217. doi: 10.1371/journal.pone.0035217.
- Ebadi, M., Bashir, R. M., Heidrick, M. L., Hamada, F. M., Refaey, H. E., Hamed, A., & Lassi, N. K. (1997). Neurotrophins and their receptors in nerve injury and repair. *Neurochemistry International*, *30*(4-5), 347-374.

- Elenkov, I. J., & Chrousos, G. P. (1999). Stress Hormones, Th1/Th2 patterns, Pro/Anti-inflammatory Cytokines and Susceptibility to Disease. *Trends in Endocrinology and Metabolism*, *10*(9), 359-368.
- Erickson, K. I., Leckie, R. L., & Weinstein, A. M. (2014). Physical activity, fitness, and gray matter volume. *Neurobiology of Aging*, *35* Suppl 2, S20-28. doi: 10.1016/j.neurobiolaging.2014.03.034.
- Erickson, K. I., Prakash, R. S., Voss, M. W., Chaddock, L., Heo, S., McLaren, M., & Kramer, A. F. (2010). Brain-derived neurotrophic factor is associated with age-related decline in hippocampal volume. *The Journal of Neuroscience: the Official Journal of the Society for Neuroscience*, *30*(15), 5368-5375. doi: 10.1523/JNEUROSCI.6251-09.2010.
- Erickson, K. I., Prakash, R. S., Voss, M. W., Chaddock, L., Hu, L., Morris, K. S., & Kramer, A. F. (2009). Aerobic fitness is associated with hippocampal volume in elderly humans. *Hippocampus*, *19*(10), 1030-1039. doi: 10.1002/hipo.20547.
- Erickson, K. I., Raji, C. A., Lopez, O. L., Becker, J. T., Rosano, C., Newman, A. B., & Kuller, L. H. (2010). Physical activity predicts gray matter volume in late adulthood: the Cardiovascular Health Study. *Neurology*, *75*(16), 1415-1422. doi: 10.1212/WNL.0b013e3181f88359.
- Erickson, K. I., Voss, M. W., Prakash, R. S., Basak, C., Szabo, A., Chaddock, L., & Kramer, A. F. (2011). Exercise training increases size of hippocampus and improves memory. *Proceedings of the National Academy of Sciences of the United States of America*, *108*(7), 3017-3022. doi: 10.1073/pnas.1015950108.
- Fang, Z. H., Lee, C. H., Seo, M. K., Cho, H., Lee, J. G., Lee, B. J., & Kim, Y. H. (2013). Effect of treadmill exercise on the BDNF-mediated pathway in the hippocampus of stressed rats. *Neuroscience Research*, *76*(4), 187-194. doi: 10.1016/j.neures.2013.04.005.
- Fernandez, O., Fernandez, V., Guerrero, M., Leon, A., Lopez-Madrona, J. C., Alonso, A., & de Ramon, E. (2012). Multiple sclerosis prevalence in Malaga, Southern Spain estimated by the capture-recapture method. *Multiple Sclerosis*, *18*(3), 372-376. doi: 10.1177/1352458511421917.
- Ferrer, I., Goutan, E., Marin, C., Rey, M. J., & Ribalta, T. (2000). Brain-derived neurotrophic factor in Huntington disease. *Brain Research*, *866*(1-2), 257-261.
- Ferris, L. T., Williams, J. S., & Shen, C. L. (2007). The effect of acute exercise on serum brain-derived neurotrophic factor levels and cognitive function. *Medicine and Science in Sports and Exercise*, *39*(4), 728-734. doi: 10.1249/mss.0b013e31802f04c7.
- Feys, P., Moundjian, L., Van Halewyck, F., Wens, I., Eijnde, B. O., Van Wijmeersch, B., & Van Asch, P. (2017). Effects of an individual 12-week community-

- located "start-to-run" program on physical capacity, walking, fatigue, cognitive function, brain volumes, and structures in persons with multiple sclerosis. *Multiple Sclerosis*, 1352458517740211. doi: 10.1177/1352458517740211.
- Filippini, G., Del Giovane, C., Vacchi, L., D'Amico, R., Di Pietrantonj, C., Beecher, D., & Salanti, G. (2013). Immunomodulators and immunosuppressants for multiple sclerosis: a network meta-analysis. *The Cochrane Database of Systematic Reviews*(6), CD008933. doi: 10.1002/14651858.CD008933.pub2.
- Fisniku, L. K., Chard, D. T., Jackson, J. S., Anderson, V. M., Altmann, D. R., Miszkiel, K. A., & Miller, D. H. (2008). Gray matter atrophy is related to long-term disability in multiple sclerosis. *Annals of Neurology*, 64(3), 247-254. doi: 10.1002/ana.21423.
- Floel, A., Ruscheweyh, R., Kruger, K., Willemer, C., Winter, B., Volker, K., & Knecht, S. (2010). Physical activity and memory functions: are neurotrophins and cerebral gray matter volume the missing link? *NeuroImage*, 49(3), 2756-2763. doi: 10.1016/j.neuroimage.2009.10.043.
- Frota, E. R., Rodrigues, D. H., Donadi, E. A., Brum, D. G., Maciel, D. R., & Teixeira, A. L. (2009). Increased plasma levels of brain derived neurotrophic factor (BDNF) after multiple sclerosis relapse. *Neuroscience Letters*, 460(2), 130-132. doi: 10.1016/j.neulet.2009.05.057.
- Gandhi, R., Laroni, A., & Weiner, H. L. (2010). Role of the innate immune system in the pathogenesis of multiple sclerosis. *Journal of Neuroimmunology*, 221(1-2), 7-14. doi: 10.1016/j.jneuroim.2009.10.015.
- Garg, N., & Smith, T. W. (2015). An update on immunopathogenesis, diagnosis, and treatment of multiple sclerosis. *Brain and Behavior*, 5(9), e00362. doi: 10.1002/brb3.362.
- Giesser, B. S. (2015). Exercise in the management of persons with multiple sclerosis. *Therapeutic Advances in Neurological Disorders*, 8(3), 123-130. doi: 10.1177/1756285615576663.
- Gleeson, M. (2007). Immune function in sport and exercise. *Journal of Applied Physiology*, 103(2), 693-699. doi: 10.1152/jappphysiol.00008.2007.
- Gleeson, M., Bishop, N. C., Stensel, D. J., Lindley, M. R., Mastana, S. S., & Nimmo, M. A. (2011). The anti-inflammatory effects of exercise: mechanisms and implications for the prevention and treatment of disease. *Nature Reviews. Immunology*, 11(9), 607-615. doi: 10.1038/nri3041.
- Gokhale, R., Chandrashekar, S., & Vasanthakumar, K. C. (2007). Cytokine response to strenuous exercise in athletes and non-athletes--an adaptive response. *Cytokine*, 40(2), 123-127. doi: 10.1016/j.cyto.2007.08.006.
- Golzari, Z., Shabkhiz, F., Soudi, S., Kordi, M. R., & Hashemi, S. M. (2010). Combined exercise training reduces IFN-gamma and IL-17 levels in the plasma and the supernatant of peripheral blood mononuclear cells in

- women with multiple sclerosis. *International Immunopharmacology*, 10(11), 1415-1419. doi: 10.1016/j.intimp.2010.08.008.
- Gustafsson, G., Lira, C. M., Johansson, J., Wisen, A., Wohlfart, B., Ekman, R., & Westrin, A. (2009). The acute response of plasma brain-derived neurotrophic factor as a result of exercise in major depressive disorder. *Psychiatry Research*, 169(3), 244-248. doi: 10.1016/j.psychres.2008.06.030.
- Gustafsson, S., Liang, W., & Hilke, S. (2011). Effects of voluntary running in the female mice lateral septum on BDNF and corticotropin-releasing factor receptor 2. *International Journal of Peptides*, 2011, 932361. doi: 10.1155/2011/932361.
- Halabchi, F., Alizadeh, Z., Sahraian, M. A., & Abolhasani, M. (2017). Exercise prescription for patients with multiple sclerosis; potential benefits and practical recommendations. *BMC Neurology*, 17(1), 185. doi: 10.1186/s12883-017-0960-9.
- Hardmeier, M., Wagenpfeil, S., Freitag, P., Fisher, E., Rudick, R. A., Kooijmans, M., & European, I. F. N. a. i. R. M. S. D. C. T. S. G. (2005). Rate of brain atrophy in relapsing MS decreases during treatment with IFNbeta-1a. *Neurology*, 64(2), 236-240. doi: 10.1212/01.WNL.0000149516.30155.B8.
- Heine, M., van de Port, I., Rietberg, M. B., van Wegen, E. E., & Kwakkel, G. (2015). Exercise therapy for fatigue in multiple sclerosis. *The Cochrane Database of Systematic Reviews*(9), CD009956. doi: 10.1002/14651858.CD009956.pub2.
- Hirsch, M. A., van Wegen, E. E. H., Newman, M. A., & Heyn, P. C. (2018). Exercise-induced increase in brain-derived neurotrophic factor in human Parkinson's disease: a systematic review and meta-analysis. *Translational Neurodegeneration*, 7, 7. doi: 10.1186/s40035-018-0112-1.
- Hohlfeld, R. (1997). Biotechnological agents for the immunotherapy of multiple sclerosis. Principles, problems and perspectives. *Brain*, 120.5, 865-916.
- Howells, D. W., Porritt, M. J., Wong, J. Y., Batchelor, P. E., Kalnins, R., Hughes, A. J., & Donnan, G. A. (2000). Reduced BDNF mRNA expression in the Parkinson's disease substantia nigra. *Experimental Neurology*, 166(1), 127-135. doi: 10.1006/exnr.2000.7483.
- Huang, E. J., & Reichardt, L. F. (2001). Neurotrophins: roles in neuronal development and function. *Annual Review of Neuroscience*, 24, 677-736. doi: 10.1146/annurev.neuro.24.1.677.
- Kebir, H., Kreymborg, K., Ifergan, I., Dodelet-Devillers, A., Cayrol, R., Bernard, M., & Prat, A. (2007). Human TH17 lymphocytes promote blood-brain barrier disruption and central nervous system inflammation. *Nature Medicine*, 13(10), 1173-1175. doi: 10.1038/nm1651.
- Kelamangalath, L., & Smith, G. M. (2013). Neurotrophin treatment to promote regeneration after traumatic CNS injury. *Frontiers in Biology*, 8(5), 486-495. doi: 10.1007/s11515-013-1269-8.

- Keller, C., Steensberg, A., Pilegaard, H., Osada, T., Saltin, B., Pedersen, B. K., & Neufer, P. D. (2001). Transcriptional activation of the IL-6 gene in human contracting skeletal muscle: influence of muscle glycogen content. *FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology*, 15(14), 2748-2750. doi: 10.1096/fj.01-0507fje.
- Kerschensteiner, M., Gallmeier, E., Behrens, L., Leal, V. V., Misgeld, T., Klinkert, W. E., & Hohlfeld, R. (1999). Activated human T cells, B cells, and monocytes produce brain-derived neurotrophic factor in vitro and in inflammatory brain lesions: a neuroprotective role of inflammation? *The Journal of Experimental Medicine*, 189(5), 865-870.
- Kierkegaard, M., Lundberg, I. E., Olsson, T., Johansson, S., Ygberg, S., Opava, C., & Piehl, F. (2016). High-intensity resistance training in multiple sclerosis - An exploratory study of effects on immune markers in blood and cerebrospinal fluid, and on mood, fatigue, health-related quality of life, muscle strength, walking and cognition. *Journal of the Neurological Sciences*, 362, 251-257. doi: 10.1016/j.jns.2016.01.063.
- Kjohede, T., Siemonsen, S., Wenzel, D., Stellmann, J. P., Ringgaard, S., Pedersen, B. G., & Dalgas, U. (2018). Can resistance training impact MRI outcomes in relapsing-remitting multiple sclerosis? *Multiple Sclerosis*, 24(10), 1356-1365. doi: 10.1177/1352458517722645.
- Klaren, R. E., Hubbard, E. A., Motl, R. W., Pilutti, L. A., Wetter, N. C., & Sutton, B. P. (2015). Objectively Measured Physical Activity Is Associated with Brain Volumetric Measurements in Multiple Sclerosis. *Behavioural Neurology*, 2015, 482536. doi: 10.1155/2015/482536.
- Knaepen, K., Goekint, M., Heyman, E. M., & Meeusen, R. (2010). Neuroplasticity - exercise-induced response of peripheral brain-derived neurotrophic factor: a systematic review of experimental studies in human subjects. *Sports Medicine*, 40(9), 765-801. doi: 10.2165/11534530-000000000-00000.
- Koch-Henriksen, N., & Sorensen, P. S. (2010). The changing demographic pattern of multiple sclerosis epidemiology. *Lancet Neurol*, 9(5), 520-532. doi: 10.1016/S1474-4422(10)70064-8.
- Koo, J. H., Kwon, I. S., Kang, E. B., Lee, C. K., Lee, N. H., Kwon, M. G., & Cho, J. Y. (2013). Neuroprotective effects of treadmill exercise on BDNF and PI3-K/Akt signaling pathway in the cortex of transgenic mice model of Alzheimer's disease. *Journal of Exercise Nutrition & Biochemistry*, 17(4), 151-160. doi: 10.5717/jenb.2013.17.4.151.
- Kraszula, L., Jasinska, A., Eusebio, M., Kuna, P., Glabinski, A., & Pietruczuk, M. (2012). Evaluation of the relationship between leptin, resistin, adiponectin and natural regulatory T cells in relapsing-remitting multiple sclerosis. *Neurologia i Neurochirurgia Polska*, 46(1), 22-28.

- Ksiazek-Winiarek, D. J., Szpakowski, P., & Glabinski, A. (2015). Neural Plasticity in Multiple Sclerosis: The Functional and Molecular Background. *Neural Plasticity, 2015*, 307175. doi: 10.1155/2015/307175.
- Kutzelnigg, A., Lucchinetti, C. F., Stadelmann, C., Bruck, W., Rauschka, H., Bergmann, M., & Lassmann, H. (2005). Cortical demyelination and diffuse white matter injury in multiple sclerosis. *Brain, 128*(Pt 11), 2705-2712. doi: 10.1093/brain/awh641.
- Lancaster, G. I., Halson, S. L., Khan, Q., Drysdale, P., Wallace, F., Jeukendrup, A. E., & Gleeson, M. (2004). Effects of acute exhaustive exercise and chronic exercise training on type 1 and type 2 T lymphocytes. *Exercise Immunology Review, 10*, 91-106.
- Latimer-Cheung, A. E., Pilutti, L. A., Hicks, A. L., Martin Ginis, K. A., Fenuta, A. M., MacKibbin, K. A., & Motl, R. W. (2013). Effects of exercise training on fitness, mobility, fatigue, and health-related quality of life among adults with multiple sclerosis: a systematic review to inform guideline development. *Archives of Physical Medicine and Rehabilitation, 94*(9), 1800-1828. doi: 10.1016/j.apmr.2013.04.020.
- Leal, L. G., Lopes, M. A., & Batista, M. L., Jr. (2018). Physical Exercise-Induced Myokines and Muscle-Adipose Tissue Crosstalk: A Review of Current Knowledge and the Implications for Health and Metabolic Diseases. *Frontiers in Physiology, 9*, 1307. doi: 10.3389/fphys.2018.01307.
- Leavitt, V. M., Ciriigliaro, C., Cohen, A., Farag, A., Brooks, M., Wecht, J. M., & Sumowski, J. F. (2014). Aerobic exercise increases hippocampal volume and improves memory in multiple sclerosis: preliminary findings. *Neurocase, 20*(6), 695-697. doi: 10.1080/13554794.2013.841951.
- Lemus, H. N., Warrington, A. E., & Rodriguez, M. (2018). Multiple Sclerosis: Mechanisms of Disease and Strategies for Myelin and Axonal Repair. *Neurologic Clinics, 36*(1), 1-11. doi: 10.1016/j.ncl.2017.08.002.
- Linker, R., Gold, R., & Luhder, F. (2009). Function of neurotrophic factors beyond the nervous system: inflammation and autoimmune demyelination. *Critical Reviews in Immunology, 29*(1), 43-68.
- Liu, Y. F., Chen, H. I., Wu, C. L., Kuo, Y. M., Yu, L., Huang, A. M., & Jen, C. J. (2009). Differential effects of treadmill running and wheel running on spatial or aversive learning and memory: roles of amygdalar brain-derived neurotrophic factor and synaptotagmin I. *The Journal of Physiology, 587*(Pt 13), 3221-3231. doi: 10.1113/jphysiol.2009.173088.
- Loma, I., & Heyman, R. (2011). Multiple sclerosis: pathogenesis and treatment. *Current Neuropharmacology, 9*(3), 409-416. doi: 10.2174/157015911796557911.
- Lucchinetti, C. F., Popescu, B. F., Bunyan, R. F., Moll, N. M., Roemer, S. F., Lassmann, H., & Ransohoff, R. M. (2011). Inflammatory cortical

- demyelination in early multiple sclerosis. *The New England Journal of Medicine*, 365(23), 2188-2197. doi: 10.1056/NEJMoa1100648.
- Luhder, F., Gold, R., Flugel, A., & Linker, R. A. (2013). Brain-derived neurotrophic factor in neuroimmunology: lessons learned from multiple sclerosis patients and experimental autoimmune encephalomyelitis models. *Archivum Immunologiae et Therapiae Experimentalis*, 61(2), 95-105. doi: 10.1007/s00005-012-0211-0.
- Manna, A., Piras, F., Caltagirone, C., Bossu, P., Sensi, S. L., & Spalletta, G. (2015). Left hippocampus-amygdala complex macro- and microstructural variation is associated with BDNF plasma levels in healthy elderly individuals. *Brain and Behavior*, 5(7), e00334. doi: 10.1002/brb3.334.
- Marais, L., Stein, D. J., & Daniels, W. M. (2009). Exercise increases BDNF levels in the striatum and decreases depressive-like behavior in chronically stressed rats. *Metabolic Brain Disease*, 24(4), 587-597. doi: 10.1007/s11011-009-9157-2.
- Matarese, G., Procaccini, C., & De Rosa, V. (2008). The intricate interface between immune and metabolic regulation: a role for leptin in the pathogenesis of multiple sclerosis? *Journal of Leukocyte Biology*, 84(4), 893-899. doi: 10.1189/jlb.0108022.
- Minagar, A., & Alexander, J. S. (2003). Blood-brain barrier disruption in multiple sclerosis. *Multiple sclerosis*, 9(6), 540-549. doi: 10.1191/1352458503ms965oa.
- Minty, A., Chalou, P., Derocq, J. M., Dumont, X., Guillemot, J. C., Kaghad, M., & Caput, D. (1993). Interleukin-13 is a new human lymphokine regulating inflammatory and immune responses. *Nature*, 362(6417), 248-250. doi: 10.1038/362248a0.
- Mokhtarzade, M., Motl, R., Negaresh, R., Zimmer, P., Khodadoost, M., Baker, J. S., & Ranjbar, R. (2018). Exercise-induced changes in neurotrophic factors and markers of blood-brain barrier permeability are moderated by weight status in multiple sclerosis. *Neuropeptides*, 70, 93-100. doi: 10.1016/j.npep.2018.05.010.
- Mokhtarzade, M., Ranjbar, R., Majdinasab, N., Patel, D., & Molanouri Shamsi, M. (2017). Effect of aerobic interval training on serum IL-10, TNFalpha, and adipokines levels in women with multiple sclerosis: possible relations with fatigue and quality of life. *Endocrine*, 57(2), 262-271. doi: 10.1007/s12020-017-1337-y.
- Motl, R. W., McAuley, E., & Snook, E. M. (2005). Physical activity and multiple sclerosis: a meta-analysis. *Multiple Sclerosis*, 11(4), 459-463. doi: 10.1191/1352458505ms1188oa.

- Motl, R. W., & Pilutti, L. A. (2012). The benefits of exercise training in multiple sclerosis. *Nature Reviews. Neurology*, 8(9), 487-497. doi: 10.1038/nrneuro.2012.136.
- Motl, R. W., & Sandroff, B. M. (2015). Benefits of exercise training in multiple sclerosis. *Current Neurology and Neuroscience Reports*, 15(9), 62. doi: 10.1007/s11910-015-0585-6.
- Motl, R. W., Sandroff, B. M., Kwakkel, G., Dalgas, U., Feinstein, A., Heesen, C., & Thompson, A. J. (2017). Exercise in patients with multiple sclerosis. *Lancet Neurology*, 16(10), 848-856. doi: 10.1016/S1474-4422(17)30281-8.
- Narayana, P. A., Govindarajan, K. A., Goel, P., Datta, S., Lincoln, J. A., Cofield, S. S., & The CombiRx Investigators, G. (2012). Regional cortical thickness in relapsing remitting multiple sclerosis: A multi-center study. *NeuroImage. Clinical*, 2, 120-131. doi: 10.1016/j.nicl.2012.11.009.
- Nave, K. A., & Trapp, B. D. (2008). Axon-glia signaling and the glial support of axon function. *Annual Review of Neuroscience*, 31, 535-561. doi: 10.1146/annurev.neuro.30.051606.094309.
- Neeper, S. A., Gomez-Pinilla, F., Choi, J., & Cotman, C. W. (1996). Physical activity increases mRNA for brain-derived neurotrophic factor and nerve growth factor in rat brain. *Brain Research*, 726(1-2), 49-56.
- Negaraesh, R., Motl, R. W., Mokhtarzade, M., Dalgas, U., Patel, D., Shamsi, M. M., & Baker, J. S. (2018). Effects of exercise training on cytokines and adipokines in multiple Sclerosis: A systematic review. *Multiple Sclerosis and Related Disorders*, 24, 91-100. doi: 10.1016/j.msard.2018.06.008.
- Nortvedt, M. W., Riise, T., & Maeland, J. G. (2005). Multiple sclerosis and lifestyle factors: the Hordaland Health Study. *Neurological Science*, 26(5), 334-339. doi: 10.1007/s10072-005-0498-2.
- O'Connor, K. C., Bar-Or, A., & Hafler, D. A. (2001). The neuroimmunology of multiple sclerosis: possible roles of T and B lymphocytes in immunopathogenesis. *Journal of Clinical Immunology*, 21(2), 81-92.
- Olsson, T. (1995). Critical influences of the cytokine orchestration on the outcome of myelin antigen-specific T-cell autoimmunity in experimental autoimmune encephalomyelitis and multiple sclerosis. *Immunological Reviews*, 144, 245-268.
- Opara, J. A., Broła, W., Wylegala, A. A., & Wylegala, E. (2016). Uhthoff's phenomenon 125 years later - what do we know today? *Journal of Medicine and Life*, 9(1), 101-105.
- Orton, S. M., Herrera, B. M., Yee, I. M., Valdar, W., Ramagopalan, S. V., Sadovnick, A. D., & Canadian Collaborative Study, G. (2006). Sex ratio of multiple sclerosis in Canada: a longitudinal study. *Lancet Neurology*, 5(11), 932-936. doi: 10.1016/S1474-4422(06)70581-6.

- Ozenci, V., Kouwenhoven, M., & Link, H. (2002). Cytokines in multiple sclerosis: methodological aspects and pathogenic implications. *Multiple Sclerosis*, *8*(5), 396-404. doi: 10.1191/1352458502ms837rr.
- Pan, W., Banks, W. A., Fasold, M. B., Bluth, J., & Kastin, A. J. (1998). Transport of brain-derived neurotrophic factor across the blood-brain barrier. *Neuropharmacology*, *37*(12), 1553-1561.
- Pearson, M. J., Mungovan, S. F., & Smart, N. A. (2018). Effect of aerobic and resistance training on inflammatory markers in heart failure patients: systematic review and meta-analysis. *Heart Failure Reviews*, *23*(2), 209-223. doi: 10.1007/s10741-018-9677-0.
- Pedersen, B. K. (2017). Anti-inflammatory effects of exercise: role in diabetes and cardiovascular disease. *European journal of clinical investigation*, *47*(8), 600-611. doi: 10.1111/eci.12781.
- Pedersen, B. K., & Febbraio, M. (2005). Muscle-derived interleukin-6--a possible link between skeletal muscle, adipose tissue, liver, and brain. *Brain, Behavior, and Immunity*, *19*(5), 371-376. doi: 10.1016/j.bbi.2005.04.008.
- Pedersen, B. K., & Hoffman-Goetz, L. (2000). Exercise and the immune system: regulation, integration, and adaptation. *Physiological Reviews*, *80*(3), 1055-1081. doi: 10.1152/physrev.2000.80.3.1055.
- Petersen, A. M., & Pedersen, B. K. (2005). The anti-inflammatory effect of exercise. *Journal of Applied Physiology*, *98*(4), 1154-1162. doi: 10.1152/jappphysiol.00164.2004.
- Pilutti, L. A., Greenlee, T. A., Motl, R. W., Nickrent, M. S., & Petruzzello, S. J. (2013). Effects of exercise training on fatigue in multiple sclerosis: a meta-analysis. *Psychosomatic Medicine*, *75*(6), 575-580. doi: 10.1097/PSY.0b013e31829b4525.
- Pilutti, L. A., Platta, M. E., Motl, R. W., & Latimer-Cheung, A. E. (2014). The safety of exercise training in multiple sclerosis: a systematic review. *Journal of the Neurological Science*, *343*(1-2), 3-7. doi: 10.1016/j.jns.2014.05.016.
- Pilli, D., Zou, A., Tea, F., Dale, R. C., & Brilot, F. (2017). Expanding Role of T Cells in Human Autoimmune Diseases of the Central Nervous System. *Frontiers in Immunology*, *8*, 652. doi: 10.3389/fimmu.2017.00652.
- Platta, M. E., Ensari, I., Motl, R. W., & Pilutti, L. A. (2016). Effect of Exercise Training on Fitness in Multiple Sclerosis: A Meta-Analysis. *Archives of Physical Medicine and Rehabilitation*, *97*(9), 1564-1572. doi: 10.1016/j.apmr.2016.01.023.
- Prakash, R. S., Snook, E. M., Motl, R. W., & Kramer, A. F. (2010). Aerobic fitness is associated with gray matter volume and white matter integrity in multiple sclerosis. *Brain Research*, *1341*, 41-51. doi: 10.1016/j.brainres.2009.06.063.
- Rasmussen, P., Brassard, P., Adser, H., Pedersen, M. V., Leick, L., Hart, E., & Pilegaard, H. (2009). Evidence for a release of brain-derived neurotrophic

- factor from the brain during exercise. *Experimental Physiology*, 94(10), 1062-1069. doi: 10.1113/expphysiol.2009.048512.
- Rivera-Quinones, C., McGavern, D., Schmelzer, J. D., Hunter, S. F., Low, P. A., & Rodriguez, M. (1998). Absence of neurological deficits following extensive demyelination in a class I-deficient murine model of multiple sclerosis. *Nature Medicine*, 4(2), 187-193.
- Rojas Vega, S., Struder, H. K., Vera Wahrmann, B., Schmidt, A., Bloch, W., & Hollmann, W. (2006). Acute BDNF and cortisol response to low intensity exercise and following ramp incremental exercise to exhaustion in humans. *Brain Research*, 1121(1), 59-65. doi: 10.1016/j.brainres.2006.08.105.
- Rudick, R. A., Fisher, E., Lee, J. C., Simon, J., & Jacobs, L. (1999). Use of the brain parenchymal fraction to measure whole brain atrophy in relapsing-remitting MS. Multiple Sclerosis Collaborative Research Group. *Neurology*, 53(8), 1698-1704.
- Ruscheweyh, R., Willemer, C., Kruger, K., Duning, T., Warnecke, T., Sommer, J., & Floel, A. (2011). Physical activity and memory functions: an interventional study. *Neurobiology of Aging*, 32(7), 1304-1319. doi: 10.1016/j.neurobiolaging.2009.08.001.
- Sacco, R., Bisecco, A., Corbo, D., Della Corte, M., d'Ambrosio, A., Docimo, R., & Bonavita, S. (2015). Cognitive impairment and memory disorders in relapsing-remitting multiple sclerosis: the role of white matter, gray matter and hippocampus. *Journal of Neurology*, 262(7), 1691-1697. doi: 10.1007/s00415-015-7763-y.
- Scalfari, A., Knappertz, V., Cutter, G., Goodin, D. S., Ashton, R., & Ebers, G. C. (2013). Mortality in patients with multiple sclerosis. *Neurology*, 81(2), 184-192. doi: 10.1212/WNL.0b013e31829a3388.
- Schwartz, C. E., Dwyer, M. G., Benedict, R., Weinstock-Guttman, B., Bergsland, N. P., Li, J., & Zivadinov, R. (2016). Reserve-related activities and MRI metrics in multiple sclerosis patients and healthy controls: an observational study. *BMC Neurology*, 16, 108. doi: 10.1186/s12883-016-0624-1.
- Sexton, C. E., Betts, J. F., Demnitz, N., Dawes, H., Ebmeier, K. P., & Johansen-Berg, H. (2016). A systematic review of MRI studies examining the relationship between physical fitness and activity and the white matter of the ageing brain. *NeuroImage*, 131, 81-90. doi: 10.1016/j.neuroimage.2015.09.071.
- Sharif, K., Watad, A., Bragazzi, N. L., Lichtbroun, M., Amital, H., & Shoenfeld, Y. (2018). Physical activity and autoimmune diseases: Get moving and manage the disease. *Autoimmunity Reviews*, 17(1), 53-72. doi: 10.1016/j.autrev.2017.11.010.
- Siffrin, V., Vogt, J., Radbruch, H., Nitsch, R., & Zipp, F. (2010). Multiple sclerosis - candidate mechanisms underlying CNS atrophy. *Trends in Neurosciences*, 33(4), 202-210. doi: 10.1016/j.tins.2010.01.002.

- Simon, J. H. (2006). Brain atrophy in multiple sclerosis: what we know and would like to know. *Multiple Sclerosis*, 12(6), 679-687. doi: 10.1177/1352458506070823.
- Stadelmann, C., Kerschensteiner, M., Misgeld, T., Bruck, W., Hohlfeld, R., & Lassmann, H. (2002). BDNF and gp145trkB in multiple sclerosis brain lesions: neuroprotective interactions between immune and neuronal cells? *Brain*, 125(Pt 1), 75-85.
- Steensberg, A., Toft, A. D., Bruunsgaard, H., Sandmand, M., Halkjaer-Kristensen, J., & Pedersen, B. K. (2001). Strenuous exercise decreases the percentage of type 1 T cells in the circulation. *Journal of Applied Physiology*, 91(4), 1708-1712. doi: 10.1152/jappl.2001.91.4.1708.
- Sutherland, G., & Andersen, M. B. (2001). Exercise and multiple sclerosis: physiological, psychological, and quality of life issues. *The Journal of Sports Medicine and Physical Fitness*, 41(4), 421-432.
- Szuhany, K. L., Bugatti, M., & Otto, M. W. (2015). A meta-analytic review of the effects of exercise on brain-derived neurotrophic factor. *Journal of Psychiatric Research*, 60, 56-64. doi: 10.1016/j.jpsychires.2014.10.003.
- Thompson, A. J., Banwell, B. L., Barkhof, F., Carroll, W. M., Coetzee, T., Comi, G., & Cohen, J. A. (2018). Diagnosis of multiple sclerosis: 2017 revisions of the McDonald criteria. *Lancet Neurology*, 17(2), 162-173. doi: 10.1016/S1474-4422(17)30470-2.
- Tongiorgi, E., Sartori, A., Baj, G., Bratina, A., Di Cola, F., Zorzon, M., & Pizzolato, G. (2012). Altered serum content of brain-derived neurotrophic factor isoforms in multiple sclerosis. *Journal of the Neurological Science*, 320(1-2), 161-165. doi: 10.1016/j.jns.2012.07.016.
- Trapp, B. D., & Nave, K. A. (2008). Multiple sclerosis: an immune or neurodegenerative disorder? *Annual Review of Neuroscience*, 31, 247-269. doi: 10.1146/annurev.neuro.30.051606.094313.
- Vacaras, V., Major, Z. Z., & Buzoianu, A. D. (2017). Brain-derived neurotrophic factor levels under chronic natalizumab treatment in multiple sclerosis. A preliminary report. *Neurologia i Neurochirurgia Polska*, 51(3), 221-226. doi: 10.1016/j.pjnns.2017.03.002.
- Verstynen, T. D., Lynch, B., Miller, D. L., Voss, M. W., Prakash, R. S., Chaddock, L., & Erickson, K. I. (2012). Caudate Nucleus Volume Mediates the Link between Cardiorespiratory Fitness and Cognitive Flexibility in Older Adults. *Journal of Aging Research*, 2012, 939285. doi: 10.1155/2012/939285.
- Vollmer, T., Signorovitch, J., Huynh, L., Galebach, P., Kelley, C., DiBernardo, A., & Sasane, R. (2015). The natural history of brain volume loss among patients with multiple sclerosis: a systematic literature review and meta-analysis. *Journal of the Neurological Science*, 357(1-2), 8-18. doi: 10.1016/j.jns.2015.07.014.

- Weinstock-Guttman, B., Zivadinov, R., Tamano-Blanco, M., Abdelrahman, N., Badgett, D., Durfee, J., & Ramanathan, M. (2007). Immune cell BDNF secretion is associated with white matter volume in multiple sclerosis. *Journal of Neuroimmunology*, *188*(1-2), 167-174. doi: 10.1016/j.jneuroim.2007.06.003.
- White, L. J., & Castellano, V. (2008). Exercise and brain health--implications for multiple sclerosis: Part 1--neuronal growth factors. *Sports Medicine*, *38*(2), 91-100. doi: 10.2165/00007256-200838020-00001.
- Wujek, J. R., Bjartmar, C., Richer, E., Ransohoff, R. M., Yu, M., Tuohy, V. K., & Trapp, B. D. (2002). Axon loss in the spinal cord determines permanent neurological disability in an animal model of multiple sclerosis. *Journal of Neuropathology and Experimental Neurology*, *61*(1), 23-32.
- Ye, X., Tai, W., & Zhang, D. (2012). The early events of Alzheimer's disease pathology: from mitochondrial dysfunction to BDNF axonal transport deficits. *Neurobiology of Aging*, *33*(6), 1122 e1121-1110. doi: 10.1016/j.neurobiolaging.2011.11.004.
- Zamvil, S. S., & Steinman, L. (2003). Diverse targets for intervention during inflammatory and neurodegenerative phases of multiple sclerosis. *Neuron*, *38*(5), 685-688.
- Zhu, J., & Paul, W. E. (2008). CD4 T cells: fates, functions, and faults. *Blood*, *112*(5), 1557-1569. doi: 10.1182/blood-2008-05-078154.
- Zuccato, C., & Cattaneo, E. (2007). Role of brain-derived neurotrophic factor in Huntington's disease. *Progress in Neurobiology*, *81*(5-6), 294-330. doi: 10.1016/j.pneurobio.2007.01.003.