

1 **Title:** Thinking small: zinc-sensing by the gut epithelium

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15 **Abbreviations:** ligand gated ion channels, LGIC; *hold on, don't rush'*, Hodor; major  
16 histocompatibility complex, MHC; target of rapamycin, TOR.

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## 18 **Main text**

19 Intestinal sensing is critical to maintain homeostasis. The gut epithelium is predominantly  
20 inhabited by enterocytes that, in addition to their barrier and absorptive function, may play a  
21 surveillance role during nutrient uptake. Even insects such as *Drosophila melanogaster* possess a  
22 sophisticated nutrient-sensing system. This suggests that enterocytes translate nutrient cues into  
23 signals that regulate the local intestinal epithelium and likely impact the entire organism (growth  
24 and development, immunity, *etc.*).

25 Miguel-Aliaga's group has recently investigated nutrient sensing by the absorptive enterocytes  
26 in *Drosophila melanogaster*.<sup>1</sup> They demonstrated that enterocyte-specific deletion of the protein  
27 transporter CG11340 (or pHCl-2) caused developmental delay in larvae. For this reason, they  
28 named it Hodor (*'hold on, don't rush'*). Hodor is a pH-sensitive, zinc-gated chloride channel that  
29 belongs to the Cys-loop subfamily of ligand-gated ion channels (LGIC). Hodor loss resulted in:  
30 brain insulin-like peptide 2 accumulation; increased autophagy; decreased food intake and lower  
31 activation of target of rapamycin (TOR); and the acidification of autophagic compartments in the  
32 copper cell region of the gut (acidic region of the middle midgut in *Drosophila* constituted by

33 copper cells and Hodor-expressing interstitial cells). These observations indicate that TOR  
34 activation in Hodor-expressing interstitial cells regulates feeding and larval growth by promoting  
35 food intake and systemic insulin signaling.<sup>1</sup>

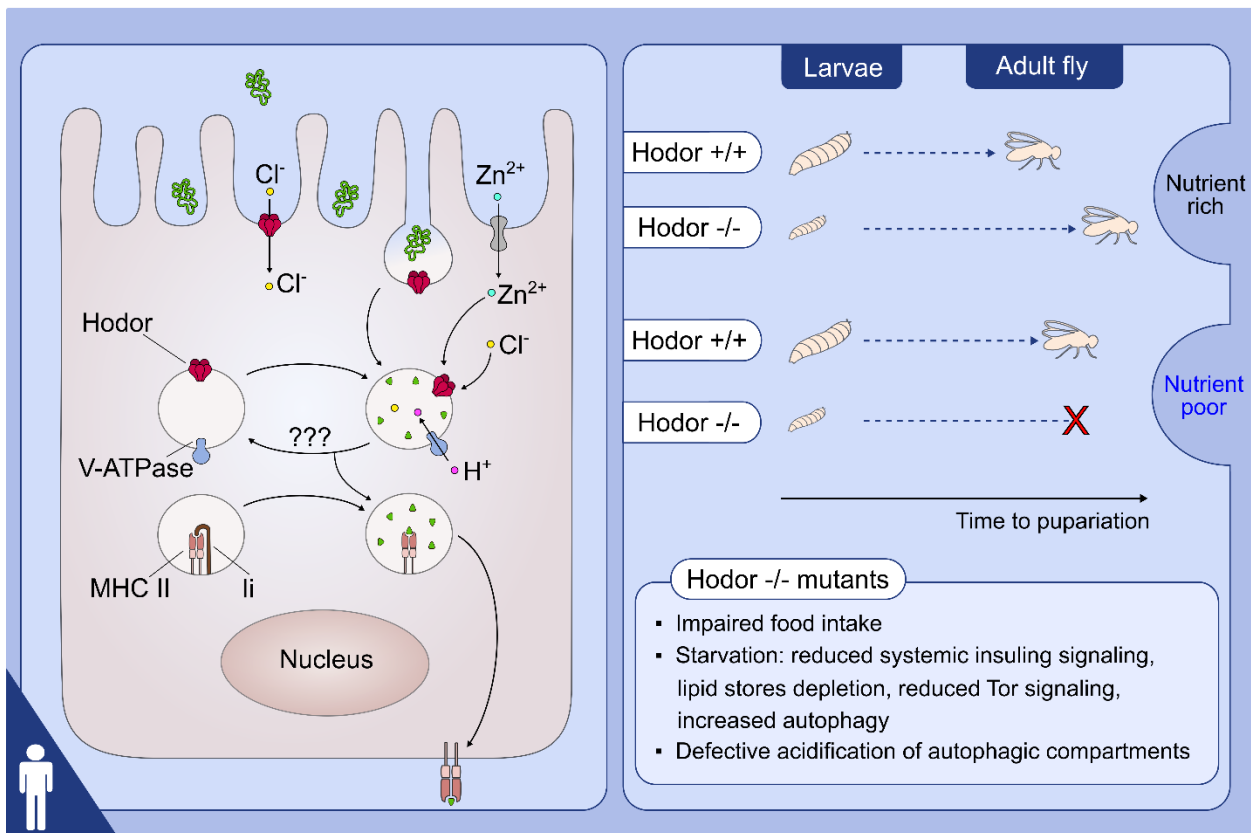
36 Initially, LGICs were thought to be neurotransmitter-gated receptors. However, this concept has  
37 been updated with the discovery of prokaryotic homologs, a zinc-activated channel<sup>2,3</sup> and the  
38 detection of LGIC in non-neuronal tissues.<sup>4,5</sup> The recent identification of Hodor as a new LGIC  
39 expressed in the gut of some invertebrates, and its activation by zinc, strengthens the notion that  
40 LGICs were incorporated by the nervous system for fast synaptic signaling.

41 Absorptive enterocytes are equipped with major histocompatibility complex (MHC) class II for  
42 presentation of extracellular antigens.<sup>6</sup> Considering that under basal conditions enterocytes  
43 express low levels of co-stimulatory molecules (*e.g.*, CD80, CD86, CD40),<sup>7</sup> oral tolerance to foods  
44 may be partly induced by epithelial-cell antigen presentation in the healthy gut. During nutrient  
45 transport, some luminal antigens are endocytosed and processed in the lysosomal compartment,  
46 binding to MHC-II in late endosomes. Miguel-Aliaga's group shows that Hodor was specifically  
47 enriched in late endosomes or lysosomes with apical localization.<sup>1</sup> For adequate antigen  
48 presentation, these antigens must be partly hydrolyzed, which requires lysosomal pH  
49 acidification for optimal protease activity. The authors found that Hodor deficiency results in  
50 defective acidification of these compartments in *Drosophila*.<sup>1</sup> Therefore, if a similar zinc-sensing  
51 mechanism was expressed in humans, zinc deficiency could alter lysosomal antigen degradation  
52 in enterocytes, potentially affecting antigen presentation.

53 Hodor-related alterations of lysosomal pH may have opposed effects in terms of antigen  
54 presentation. On the one hand, Hodor dysfunction could limit lysosomal antigen degradation,  
55 potentially reducing antigen presentation. On the other hand, it could prevent excessive antigen  
56 degradation into immune-silent small peptides, potentially increasing antigen presentation. The  
57 outcome is presumably affected by the cell type and other regulatory mechanisms. As the  
58 enterocyte is a highly degradative cell type specialized in nutrient supply, zinc shortage may  
59 protect luminal antigens from full lysosomal degradation enabling antigen presentation at the  
60 gut epithelium.

61 The role of zinc in mucosal integrity is well known, particularly as it pertains to the barrier  
 62 function of intestinal epithelial cells. For example, zinc deficiency has been reported in patients  
 63 with chronic diarrhea or inflammatory bowel disease. In this context, it is possible that the lack  
 64 of zinc (*i.e.*, reduced Hodor signaling) exacerbates pathology via increased enterocyte-driven  
 65 antigen presentation.<sup>8,9</sup> Especially considering that intestinal inflammation or exposure to certain  
 66 food antigens (*e.g.*, gliadin in celiac disease) has been shown to upregulate enterocyte expression  
 67 of MHC-II and co-stimulatory molecules.

68 The intestinal epithelium is being increasingly recognized as a multifaceted player of the  
 69 gastrointestinal tract. It plays a central role in nutrient absorption while providing a first line of  
 70 luminal surveillance that guides mucosal immune responses. The interdisciplinary investigation  
 71 of enterocyte nutrient-sensing pathways by which enterocytes communicate to other cells will  
 72 advance the understanding of gut homeostasis and immune function.



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74 **Figure 1.** Hodor was specifically enriched in the late endosomes or lysosomes with apical  
 75 localization and in the brush border of interstitial cells of *Drosophila melanogaster*. Zinc-sensing

76 by Hodor promotes chloride transport in/out cytoplasm and lysosomes. If expressed in humans,  
77 Hodor-mediated chloride transport across lysosomal membranes could sustain the activity of the  
78 proton-pumping vacuolar-type ATPase (V-ATPase) that maintains lysosomal acidity. During  
79 luminal transport of antigens by absorptive enterocytes, some endocytosed antigens can be  
80 processed in the acidified lysosomal compartment. Later these peptides bind to MHC-II in late  
81 endosomes upon invariant chain (Ii) removal.

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