

Prioritizing Phase I Treatment Options Through Preclinical Testing in Personalized Tumorgraft

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A 29 years old young woman with a history of advanced adenoid cystic carcinoma (ACC) resistant to standard of care treatments presented to our phase I clinic seeking treatment with experimental therapeutics. The patient was diagnosed with ACC 11 years before and had been treated with surgery, radiation treatment, and several lines of conventional treatments including platinum, anthracyclines, and imatinib mesylate. Eleven months before being seen in our clinic she had developed a brain metastasis that had been surgically resected. A personalized tumorgraft was successfully established from this lesion by implanting fragments of tumor materials in immunocompromised mice as described by our group.¹ At the time of enrollment in the clinical trial she had pulmonary and liver metastasis and was progressing with growth of a preexisting liver metastasis and development of a new liver lesion compared with a CT-scan performed six months before (Before Baseline CT-scan), as depicted in Figure 1. A brain MRI showed a stable 2 mm brain lesion. The patient was asymptomatic with ECOG 0 and normal liver, bone marrow and kidney functions.

In order to determine which phase I clinical studies could be more appropriate for the patient, we characterized her tumor for *KRAS* mutations and *HER2* amplification and found that

it to be *KRAS* wild type and not HER2 amplified respectively. Given that she had a personalized tumorgraft model developed from her brain metastases we utilized the model to evaluate a battery of anticancer agents, both conventional and experimental. Briefly, a tumor specimen obtained at the time of removal of her brain tumor had been transplanted and propagated in nude mice. Once it was in exponential growth phase, cohorts of mice with tumor size of 150-300 mm³ were randomized to several treatment groups. The results of these studies are listed in Figure 2. One of the most effective agents was a monoclonal antibody against the Insulin Growth Factor Receptor 1 (IGF1-R). As shown in Figure 2, intraperitoneal administration of this agent at a dose of 40mg/Kg every 3 weeks resulted in about a 76 % tumor growth inhibition compared to untreated control after 49 days of treatment. Although we did not observed tumor regression, treatment results in a significant cytostatic effect, and we considered the tumor sensible to treatment (p<0.001, t-test analysis).

At that time, we were conducting a phase I trial evaluating the combination of figitumumab, an IGF1R monoclonal antibody and PF00299804, a pan-HER inhibitor.² Based on the mouse data obtained with the IGF1R inhibitor, and on the fact that here tumor was *KRAS* wild-type, the patient was enrolled in this trial. Figitumumab was administered IV every three weeks and PF00299804 was given on a daily orally administered schedule. After four cycles of treatment with the combination of PF00299804 and figitumumab patient developed a severe diarrhea, mostly related to the pan-HER toxicity and treatment with PF00299804 was discontinued, at that time patient tumor assessment was showing stable disease. Then patient continued treatment with figitumumab alone for four more cycles. Treatment with the IGF1R inhibitor showed a good safety profile and resulted in a minor response in the rapidly growing

liver lesion that lasted until cycle 8 of treatment.. At that time, the patient progressed with a new brain lesion and was taken off study. The tumor remained controlled outside the brain.

ACC are very rare variants of adenocarcinoma that most often arise from salivary glands. Roughly 500 new cases of ACC are diagnosed in USA each year³. The natural history of the disease can be characterized either by an indolent growth in some patients or by an aggressive and rapidly progressive disease. The overall 10-years survival for patients is about 50%, but when metastases occur the median duration of survival is about 3-years⁴⁻⁶. Because of the rarity of this disease there are few clinical trials investigating systemic therapy. Data, from a recently published meta-analysis, reported activity for platinum and/or doxorubicin-based regimen⁷. Imatinib and EGFR inhibitors have been recently evaluated in this disease with limited success.^{8,9,10,11} There is no standard approach for patients who progress to conventional treatments. As such enrollment in clinical trials with novel agents is an accepted approach. Because of the paucity of molecular data in this cancer, however, selection of an appropriate trial is done, in general empirically. This situation is not limited to ACC and in fact is the norm for most solid tumors: patients are enrolled in early clinical trials mostly based on clinical and not molecular eligibility criteria. However, the failure rate in phase I clinical trials remains extremely high with most patients deriving no benefit. In fact, only in studies in which there is a clear connection between a genetic alteration and an inhibitor exist showed high response rate in phase I studies.¹²⁻¹⁴

During the last years our group has been involved in the development of personalized mouse models from patients with cancer. These models are useful for drug screening and

biomarker development and thus can be used to prioritize and rank effective agents against an individual cancer. As illustrated in this case, we were able to screen a large set of agents, both experimental and conventional against the personalized tumorgraft model. The data from these studies suggest that this cancer may be sensitive to *nab*-paclitaxel, temozolomide, and inhibitors of IGF1R, mTOR and FGFR inhibitors providing several therapeutic opportunities. Because the IGF1R inhibitor trial was available we elected to enroll the patient in that study saving the other conventional opportunities for the future. In fact, she is now been treated with temozolamide based on the results from these experiments. Overall the data supports the use of personalized tumorgraft as a standard to test experimental agents prior to administration of patients. Furthermore, the personalized tumorgraft model, with a clinically validated susceptibility to IGF1R blockade is now an interesting preclinical tool to comparatively explore other IGF1R inhibitors, design combinations and investigate biomarkers. This is, for example, illustrated in our recent report showing that PALB2 mutations is a strong candidate biomarker of response to DNA damaging agents.¹⁵

There is however some limitations that will need to be addressed before this strategy can be broadly implemented in phase I clinical studies. Patients need to have a personalized tumorgraft model established. For this patient that was accomplished by implanting excess tissue from a clinically indicated surgical resection. However most phase I candidates do not need surgery thus collecting fresh tumor tissue for the generation of a personalized tumorgraft model can be a challenge. Additionally, the development, propagation of the personalized tumorgraft model and drug testing takes 6-8 months, which is often not available for most phase I candidates. Moreover, failure rate in tumorgraft establishment and the possibility of molecular

signature discordance between two lesions from different organs need to be considered. Thus, early selection of potential candidates is critical. Finally is the selection of agents to be tested. The number of drugs in development too large and there has to be a strategy to prioritize which drugs to be tested in the models. In this sense, integration of molecular testing to rank order candidates is be critical.

The next prevailing question is to test this strategy in a clinical trial to show is feasible and beneficial. A plausible design for example would be patients with advanced cancer with accessible primary tumor and metastatic lesions from whom a personalized tumorgraft model can be generated and available at the time of second line treatment. These lesions can be profiled with a battery of biomarkers to select a set of 5-10 agents to be tested in the model to select the ideal phase I study for the patient. Obviously, there are significant logistic and feasibility issues with such an approach but the reality is that the field is stagnant with little progress and new, albeit risky, approaches are needed.

REFERENCES

1. Rubio-Viqueira B, Jimeno A, Cusatis G, et al: An in vivo platform for translational drug development in pancreatic cancer. *Clin Cancer Res* 12:4652-61, 2006
2. Engelman JA, Zejnullahu K, Gale CM, et al: PF00299804, an irreversible pan-ERBB inhibitor, is effective in lung cancer models with EGFR and ERBB2 mutations that are resistant to gefitinib. *Cancer Res* 67:11924-32, 2007

3. Renehan A, Gleave EN, Hancock BD, et al: Long-term follow-up of over 1000 patients with salivary gland tumours treated in a single centre. *Br J Surg* 83:1750-4, 1996
4. Spiro RH: Distant metastasis in adenoid cystic carcinoma of salivary origin. *Am J Surg* 174:495-8, 1997
5. Spiro RH: Management of malignant tumors of the salivary glands. *Oncology (Williston Park)* 12:671-80; discussion 683, 1998
6. Terhaard CH, Lubsen H, Van der Tweel I, et al: Salivary gland carcinoma: independent prognostic factors for locoregional control, distant metastases, and overall survival: results of the Dutch head and neck oncology cooperative group. *Head Neck* 26:681-92; discussion 692-3, 2004
7. Laurie SA, Ho AL, Fury MG, et al: Systemic therapy in the management of metastatic or locally recurrent adenoid cystic carcinoma of the salivary glands: a systematic review. *Lancet Oncol*
8. Hotte SJ, Winkquist EW, Lamont E, et al: Imatinib mesylate in patients with adenoid cystic cancers of the salivary glands expressing c-kit: a Princess Margaret Hospital phase II consortium study. *J Clin Oncol* 23:585-90, 2005
9. Pfeffer MR, Talmi Y, Catane R, et al: A phase II study of Imatinib for advanced adenoid cystic carcinoma of head and neck salivary glands. *Oral Oncol* 43:33-6, 2007
10. Locati LD, Bossi P, Perrone F, et al: Cetuximab in recurrent and/or metastatic salivary gland carcinomas: A phase II study. *Oral Oncol* 45:574-8, 2009
11. Agulnik M, Cohen EW, Cohen RB, et al: Phase II study of lapatinib in recurrent or metastatic epidermal growth factor receptor and/or erbB2 expressing adenoid cystic

carcinoma and non adenoid cystic carcinoma malignant tumors of the salivary glands. *J Clin Oncol* 25:3978-84, 2007

12. Flaherty KT, Puzanov I, Kim KB, et al: Inhibition of mutated, activated BRAF in metastatic melanoma. *N Engl J Med* 363:809-19

13. Kwak EL, Bang YJ, Camidge DR, et al: Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N Engl J Med* 363:1693-703

14. Von Hoff DD, LoRusso PM, Rudin CM, et al: Inhibition of the hedgehog pathway in advanced basal-cell carcinoma. *N Engl J Med* 361:1164-72, 2009

15. Villarroel MC, Rajeshkumar NV, Garrido-Laguna I, et al: Personalizing cancer treatment in the age of global genomic analyses: PALB2 gene mutations and the response to DNA damaging agents in pancreatic cancer. *Mol Cancer Ther* 10:3-8

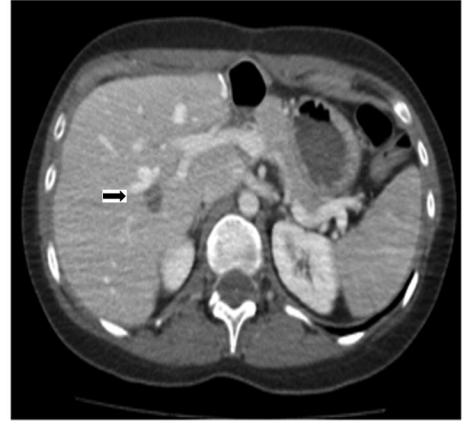
Tumor Assessment

Before Baseline

Baseline

End of Study

Liver



Brain

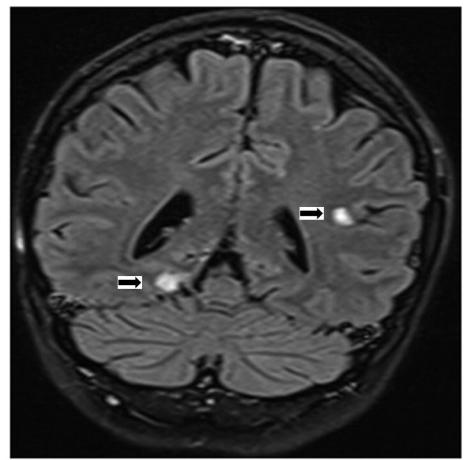
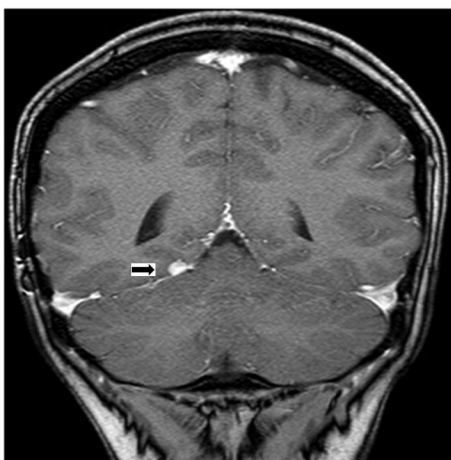
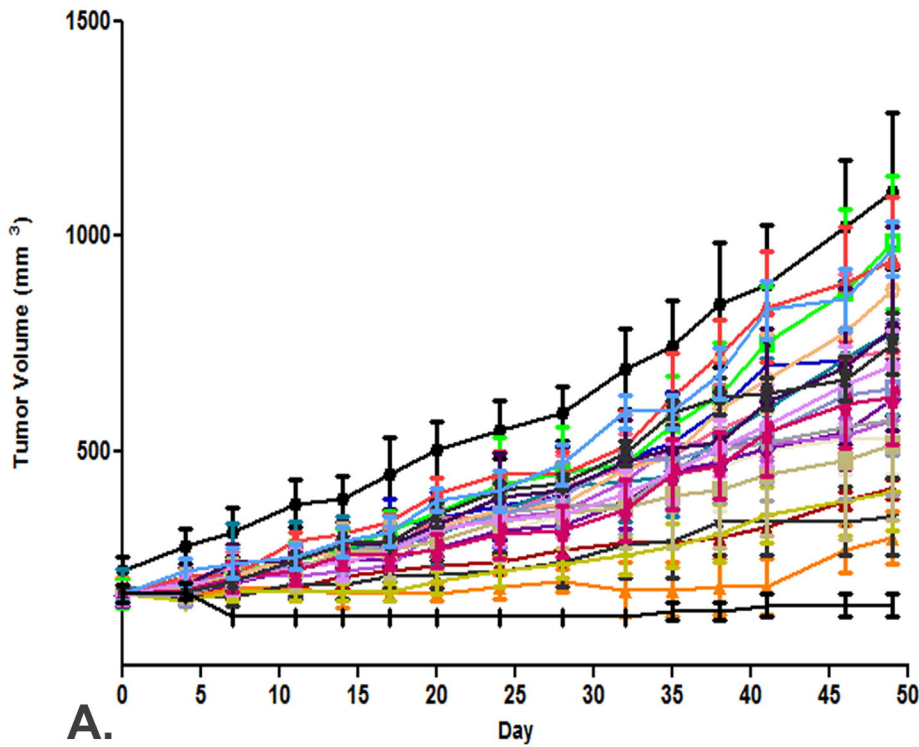


Fig 1

Patient Xenograft vs Treatment

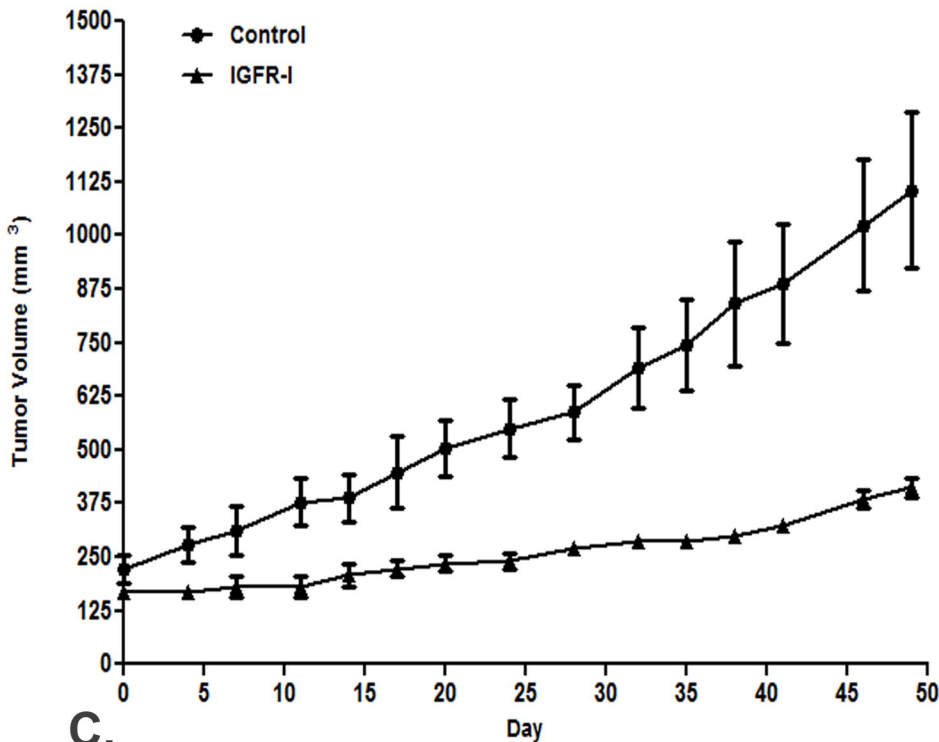


- Control
- ▲ IGFR-I
- ▲ Abraxane
- ▲ 5-Fluorouracil
- ▲ Bortezomib
- ▲ Trastuzumab
- ▲ Gemcitabine
- ▲ Ifosfamide
- ▲ Irinotecan
- ▲ Oxaliplatin
- ▲ Doxil
- ▲ FGFR-I
- ▲ Tensirolimus
- ▲ Topotecan
- ▲ Imatinib
- ▲ Dasatinib
- ▲ Lapatinib
- ▲ Sorafenib
- ▲ Sunitinib
- ▲ Vorinostat
- ▲ Vinorelbine
- ▲ Mitomicin-C
- ▲ Tomozolomide

Drug	Activity Rating (AR)%
IGFR-I	72%
Abraxane	73%
5-Fluorouracil	13%
Bortezomib	19%
Trastuzumab	12%
Gemcitabine	<0%
Ifosfamide	<0%
Irinotecan	48%
Oxaliplatin	34%
Doxil	41%
FGFR-I	74%
Tensirolimus	67%
Topotecan	41%
Tomozolomide	102%
Imatinib	<0%
Dasatinib	27%
Lapatinib	10%
Sorafenib	50%
Sunitinib	20%
Vorinostat	31%
Vinorelbine	14%
Mitomicin-C	<0%

B.

Patient Xenograft vs IGFR-Inhibitor



Gene	Status
HER2/Neu	No amplified
KRAS	WT

D.

Fig 2